Ghrelin (Human/Mouse/Rat) ELISA Kit

Catalog Number KA1863

96 assays

Version: 02

Intended for research use only

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Introduction

Intended Use

The Ghrelin (Human, Mouse, Rat) ELISA Kit is an in vitro quantitative assay for detecting Ghrelin peptide based on the principle of Competitive Enzyme Immunoassay.

Background

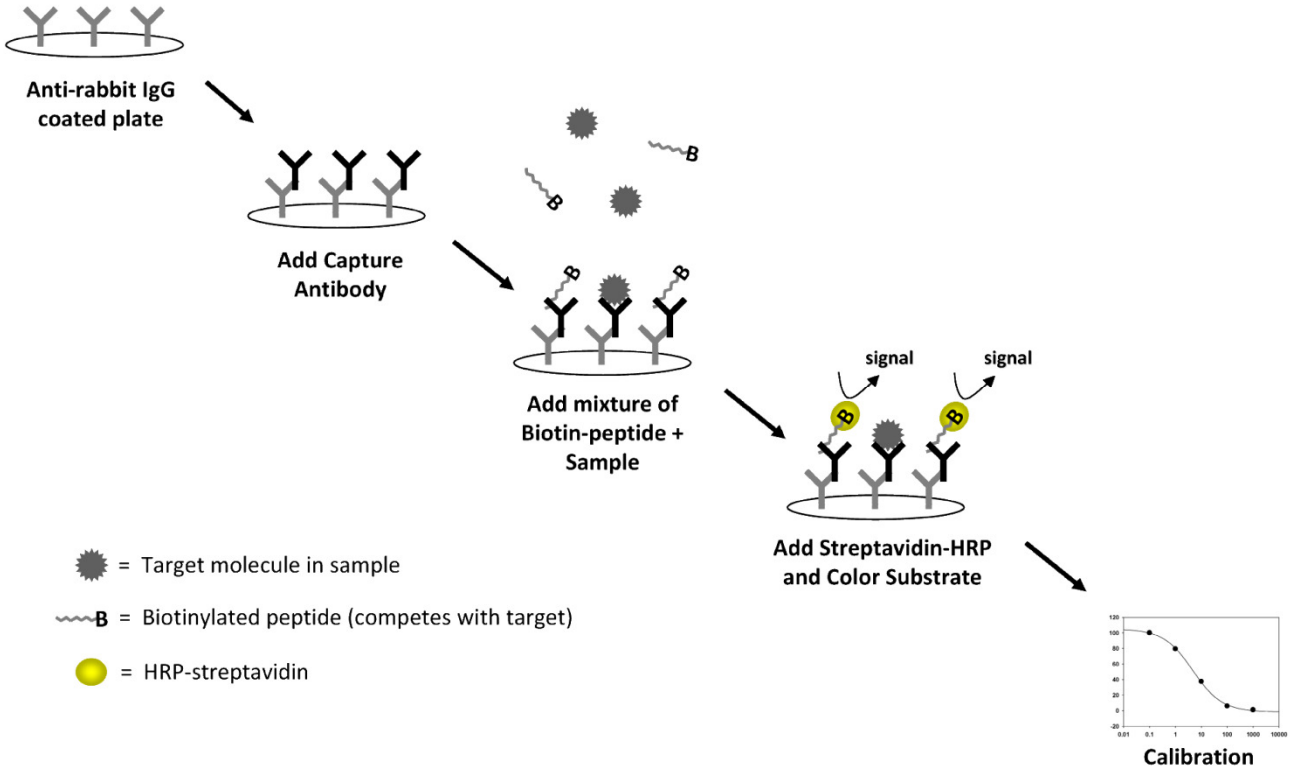
Obesity, which is characterized by excessive accumulation of adipose tissue in the body, has become one of the greatest public health challenges. Obesity is not only associated with health problems linked to increased weight-dependent pressure overload on lung, joints and bones, but also a important risk factor for life-threatening diseases such as cardiovascular diseases, type 2 diabetes and certain cancers.

Ghrelin is synthesized as a preprohormone, and then proteolytically processed to yield a 28-amino acid peptide. Synthesis of ghrelin occurs predominantly in epithelial cells lining the fundus of the stomach, with smaller amounts produced in the placenta, kidney, pituitary and hypothalamus.

Ghrelin has emerged as the first circulating hunger hormone. Ghrelin increases food intake and thus fat mass by an action exerted at the level of the hypothalamus. They activate cells in the arcuate nucleus that include the orexigenic neuropeptide Y (NPY) neurons. Ghrelin-responsiveness of these neurons is both leptin and insulin sensitive. Ghrelin also activates the mesolimbic cholinergic-dopaminergic reward link, a circuit that communicates the hedonic and reinforcing aspects of natural rewards, such as food.
Ghrelin levels in the plasma of obese individuals are lower than those in leaner individuals except in the case of Prader-Willi syndrome-induced obesity. Those suffering from the eating disorder anorexia nervosa have high plasma levels of ghrelin compared to both the constitutionally thin and normal-weight controls. These findings suggest that ghrelin plays a role in both anorexia and obesity. Ghrelin levels are also high in patients who have cancer-induced cachexia.

Principle of the Assay

The microplate in the kit is pre-coated with anti-rabbit secondary antibody. After a blocking step and incubation of the plate with anti- Ghrelin antibody, both biotinylated Ghrelin peptide and peptide standard or targeted peptide in samples interacts competitively with the Ghrelin antibody. Uncompeted (bound) biotinylated Ghrelin peptide then interacts with Streptavidin-horseradish peroxidase (SA-HRP), which catalyzes a color development reaction. The intensity of colorimetric signal is directly proportional to the amount of biotinylated peptide-SA-HRP complex and inversely proportional to the amount of Ghrelin peptide in the standard or samples. This is due to the competitive binding to Ghrelin antibody between biotinylated Ghrelin peptide and peptides in standard or samples. A standard curve of known concentration of Ghrelin peptide can be established and the concentration of Ghrelin peptide in the samples can be calculated accordingly.
Ghrelin ELISA Kit detects inactive Ghrelin (117aa), not the acylated or active forms of Ghrelin.


Anti-rabbit IgG
coated plate
Add Capture
Antibody
B
B
Add mixture of
Biotin-peptide +
Sample
signal
signal
Add Streptavidin-HRP
and Color Substrate
= Target molecule in sample
B = Biotinylated peptide (competes with target)
= HRP-streptavidin
Calibration

General Information

Materials Supplied

Component	Amount
Ghrelin Microplate (Item A): Coated with secondary antibody.	96 wells (12 strips x 8 wells)
Wash Buffer Concentrate (20x) (Item B)	25 ml
Standard Ghrelin Peptide (Item C)	2 vials, 10 µl/vial
Anti- Ghrelin polyclonal antibody (Item N)	2 vials, 5 µl/vial
Assay Diluent A (Item D): contains 0.09% sodium azide as preservative. For Standard/Sample (serum/ plasma) diluent.	30 ml
Assay Diluent B (Item E): 5x concentrated buffer. Diluent for standards and cell culture media or other sample types.	15 ml
Biotinylated Ghrelin peptide, (Item F)	2 vials, 20 µl/vial
HRP-Streptavidin concentrate (Item G): 200x concentrated HRP-conjugated Streptavidin.	600 µl
Positive control (Item M)	1 vial, 100 µl
TMB One-Step Substrate Reagent (Item H): 3, 3', 5, 5'-tetramethylbenzidine (TMB) in buffered solution.	12 ml
Stop Solution (Item I): 0.2 M sulfuric acid.	8 ml

Storage Instruction

- ✓ Standard, Biotinylated Ghrelin peptide, and Positive Control should be stored at -20℃ or -80℃ (recommended at -80℃) after arrival. Avoid multiple freeze-thaws.
- ✓ The remaining kit components may be stored at -20℃.
- ✓ Opened Microplate Wells and antibody (Item N) may be stored for up to 1 month at 2° to 8℃. Return unused wells to the pouch containing desiccant pack and reseal along entire edge.
- ✓ If stored in this manner, Abnova warranties this kit for 6 months from the date of shipment.

Materials Required but Not Supplied

- ✓ Microplate reader capable of measuring absorbance at 450 nm.
- ✓ Precision pipettes to deliver 2 µl to 1 ml volumes.
- ✓ Adjustable 1-25 ml pipettes for reagent preparation.
- ✓ 100 ml and 1 liter graduated cylinders.
- ✓ Absorbent paper.
- ✓ Distilled or deionized water.
- ✓ SigmaPlot software (or other software which can perform four-parameter logistic regression models)

- ✓ Tubes to prepare standard or sample dilutions.
- ✓ Orbital shaker
- ✓ Aluminum foil
- ✓ Saran Wrap

Assay Protocol

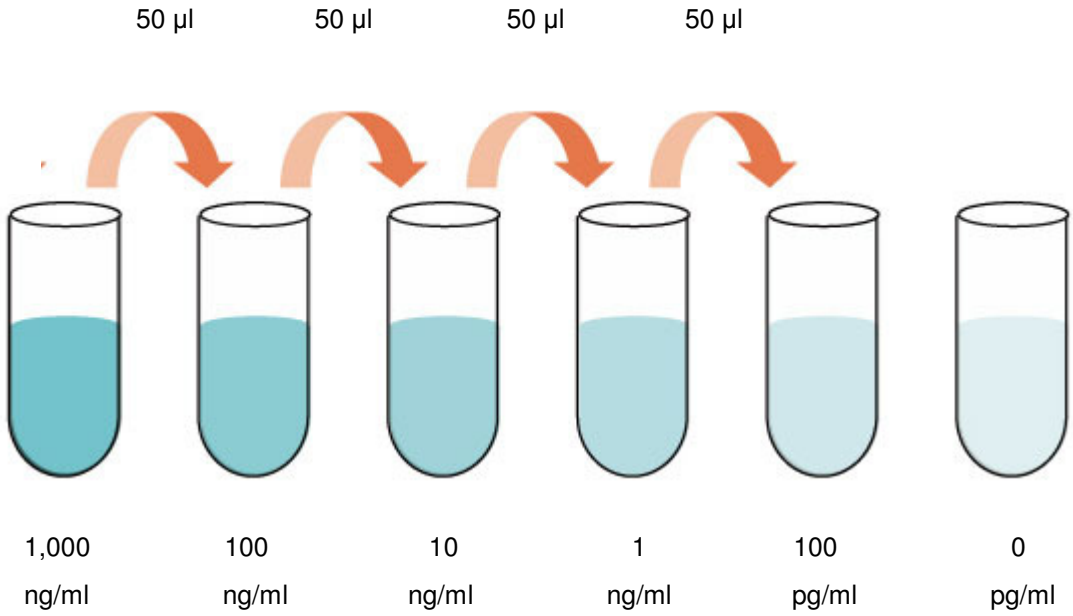
Reagent Preparation

If testing plasma or serum samples, use Assay Diluent A to dilute Item F and Item C. If testing cell culture media or other sample types, use Assay Diluent B to dilute Item F and Item C. For sample and positive control dilutions, refer to steps 6, 7, 8 and 10 of Reagent Preparation.

1. Keep kit reagents on ice during reagent preparation steps. Equilibrate plate to room temperature before opening the sealed pouch.
2. Assay Diluent B (Item E) should be diluted 5-fold with deionized or distilled water.
3. Briefly centrifuge the Anti-Ghrelin Antibody vial (Item N) before use. Add 50 µl of 1x Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently.
4. The antibody concentrate should then be diluted 100-fold with 1x Assay Diluent B. This is your anti-Ghrelin antibody working solution, which will be used in step 2 of the Assay Procedure.

   Note: the following steps may be done during the antibody incubation procedure (step 2 of Assay Procedure).
5. Briefly centrifuge the vial of Biotinylated Ghrelin (Item F) before use. Add 5 µl of Item F to 5 ml of the appropriate Assay Diluent. Pipette up and down to mix gently. The final concentration of biotinylated Ghrelin will be 10 ng/ml. This solution will only be used as the diluent in step 6 of Reagent Preparation.
6. Preparation of Standards: Label 6 microtubes with the following concentrations: 1000 ng/ml, 100 ng/ml, 10 ng/ml, 1 ng/ml, 100 pg/ml and 0 pg/ml. Pipette 450 µl of biotinylated Ghrelin solution into each tube, except for the 1000 ng/ml (leave this one empty). It is very important to make sure the concentration of biotinylated Ghrelin is 10 ng/ml in all standards.

- ✓ Briefly centrifuge the vial of Ghrelin (Item C). In the tube labeled 1000 ng/ml, pipette 8 µl of Item C and 792 µl of 10 ng/ml biotinylated Ghrelin solution (prepared in step 5 above). This is your Ghrelin stock solution (1000 ng/ml Ghrelin, 10 ng/ml biotinylated Ghrelin). Mix thoroughly. This solution serves as the first standard.
- ✓ To make the 100 ng/ml standard, pipette 50 µl of Ghrelin stock solution the tube labeled 100 ng/ml. Mix thoroughly.
- ✓ Repeat this step with each successive concentration, preparing a dilution series as shown in the illustration below. Each time, use 450 µl of biotinylated Ghrelin and 50 µl of the prior concentration until 100 pg/ml is reached. Mix each tube thoroughly before the next transfer.
- ✓ The final tube (0 pg/ml Ghrelin, 10 ng/ml biotinylated Ghrelin) serves as the zero standard (or total binding).



7. Prepare a 10-fold dilution of Item F. To do this, add 2 µl of Item F to 18 µl of the appropriate Assay Diluent. This solution will be used in steps 8 and 10.
8. Positive Control Preparation: briefly centrifuge the positive control vial (Item M). To the tube of Item M, add 101 µl 1x Assay Diluent B. Also add 2 µl of 10-fold diluted Item F (prepared in step 7) to the tube. This is a 2-fold dilution of the positive control. Mix thoroughly. The positive control is a cell culture medium sample with an expected signal between 10% and 30% of total binding (70-90% competition) if diluted as described above. It may be diluted further if desired, but be sure the final concentration of biotinylated Ghrelin is 10 ng/ml.
9. If Item B (20x Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1x Wash Buffer.
10. Sample Preparation: Use Assay Diluent A + biotinylated Ghrelin to dilute serum/plasma samples. For cell culture medium and other sample types, use 1x Assay Diluent B + biotinylated Ghrelin as the diluent. It is very important to make sure the final concentration of the biotinylated Ghrelin is 10 ng/ml in every sample.
    EXAMPLE: to make a 4-fold dilution of sample, mix together 2.5 µl of 10-fold diluted Item F (prepared in step 7), 185 µl of appropriate Assay Diluent, and 62.5 µl of your sample; mix gently. The total volume is 250 µl, enough for duplicate wells on the microplate.
    Do not use Item F diluent from Step 5 for sample preparation. If you plan to use undiluted samples, you must still add biotinylated Ghrelin to a final concentration of 10 ng/ml.
    EXAMPLE: Add 2.5 µl of 10-fold diluted Item F to 247.5 µl of sample.
    Note: Optimal sample dilution factors should be determined empirically, however you may contact us (sales@abnova.com) to obtain recommended dilution ranges for serum or plasma.
11. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use. The HRP-Streptavidin concentrate should be diluted 200-fold with 1x Assay Diluent B.
    Note: Do not use Assay Diluent A for HRP-Streptavidin preparation in Step 11.

Assay Procedure

1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100 µl anti-Ghrelin antibody (see Reagent Preparation step 4) to each well. Incubate for 1.5 hours at room temperature with gentle shaking (1-2 cycles/sec). You may also incubate overnight at 4℃.
3. Discard the solution and wash wells 4 times with 1x Wash Buffer (200-300 µl each), Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 µl of each standard (see Reagent Preparation step 6), positive control (see Reagent Preparation step 8) and sample (see Reagent Preparation step 10) into appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1-2 cycles/sec) or overnight at 4℃.
5. Discard the solution and wash 4 times as directed in Step 3.
6. Add 100 µl of prepared HRP-Streptavidin solution (see Reagent Preparation step 11) to each well. Incubate for 45 minutes at room temperature. It is recommended that incubation time should not be shorter or longer than 45 minutes.
7. Discard the solution and wash 4 times as directed in Step 3.
8. Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1-2 cycles/sec).
9. Add 50 µl of Stop Solution (Item I) to each well. Read absorbances at 450 nm immediately.

- Summary

1. Prepare all reagents, samples and standards as instructed.
2. Add 100 µl anti-Ghrelin antibody to each well. Incubate 1.5 hours at room temperature or overnight at 4℃.
3. Add 100 µl standard or sample to each well. Incubate 2.5 hours at room temperature or overnight at 4℃.
4. Add 100 µl prepared streptavidin solution. Incubate 45 minutes at room temperature.
5. Add 100 µl TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50 µl Stop Solution to each well. Read at 450 nm immediately.

Data Analysis

Calculation of Results

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit straight line through the standard points.

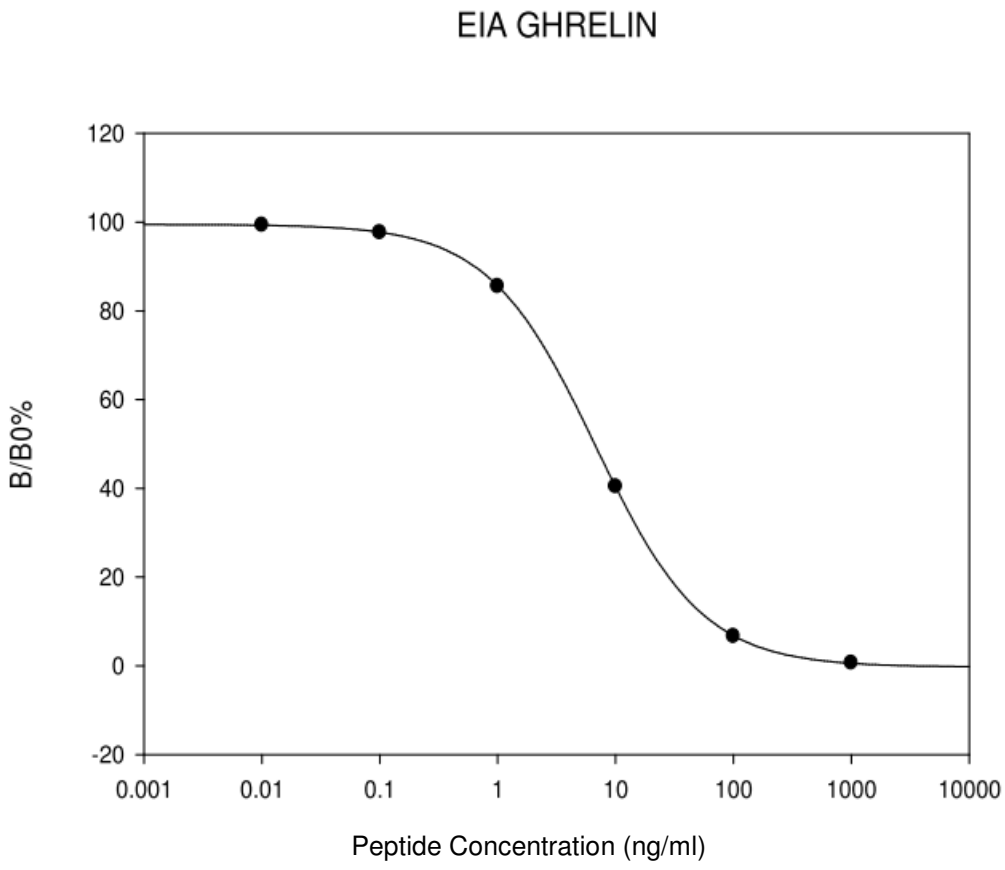
Percentage absorbance = (B – blank OD)/(Bo – blank OD) where

B = OD of sample or standard and

Bo = OD of zero standard (total binding)

Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



Performance Characteristics

- ✓ Sensitivity

  The minimum detectable concentration of Ghrelin is 161 pg/ml or 12.46pM.
- ✓ Detection Range

  0.1-1,000 ng/ml
- ✓ Reproducibility

  Intra-Assay: CV<10%

  Inter-Assay: CV<15%
- ✓ Specificity

  Cross Reactivity: This ELISA kit shows no cross-reactivity with any of the cytokines tested: Nesfatin, Angiotensin II, NPY and APC.

Resources

Troubleshooting

Problem	Cause	Solution
Poor standard curve	Inaccurate pipetting	Check pipettes
	Improper standard dilution	Ensure briefly spin the vial of Item C and dissolve the powder thoroughly by a gentle mix.
Low signal	Too brief incubation times	Ensure sufficient incubation time; assay procdure step 2 change to over night
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
Large CV	Inaccurate pipetting	Check pipettes
High background	Plate is insufficiently washed	Review the manual for proper wash. If using a plate washer, check that all ports are unobstructed.
	Contaminated wash buffer	Make fresh wash buffer
Low sensitivity	Improper storage of the ELISA kit	Store your standard at<-20℃ after reconstitution, others at 4℃. Keep substrate solution protected from light
	Stop solution	Stop solution should be added to each well before measure

References

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Plate Layout

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8								
7								
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4								
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	A	B	C	D	E	F	G	H