# ab204909 Antibody Purification Kit - Nanoparticles

# A product of Expedeon, an Abcam company

Applicable to Expedeon product codes: 260-0005, 260-0010

View ab204909 Antibody Purification Kit - Nanoparticles: <u>www.abcam.com/ab204909</u> (use <u>www.abcam.cn/ab204909</u> for China, or <u>www.abcam.co.jp/ab204909</u> for Japan)

For the purification of IgG fractions from tissue culture supernatants.

This product is for research use only and is not intended for diagnostic use.

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### 1. Overview

Antibody Purification Kit - Nanoparticles (ab204909) removes common buffer constituents found in commercial antibodies (e.g. BSA, glycine, tris and azide) that decrease the efficiency of conjugation reactions with enzymes or flurophores. ab204909 can also be used to purify antibodies from crude samples such as ascites fluid or serum.

The method involves capture of the antibody on Protein A Resin and removal of unwanted substances by a simple wash procedure, which is carried out in a standard microfuge. The purified antibody is then eluted and neutralized.

Antibodies purified using the Antibody Purification Kit - Nanoparticles are fully compatible with our Gold antibody conjugation kits, Magnetic particle conjugation kits, Latex and Europium conjugation kits (available separately).

# 2. Materials Supplied and Storage

Store kit at +4°C immediately on receipt. **Do not freeze or store the resin at room temperature.** Freezing the suspension will damage the agarose beads.

Item	Quantity		Storage
liem	1 TEST	3 TESTS	temperature
Protein A resin	1 bottle	3 bottles	+4°C
Spin Purification Columns	1 column	3 columns	+4°C
10x Binding Buffer	1 bottle	1 bottle	+4°C
Wash Buffer	1 bottle	1 bottle	+4°C
Elution Buffer	1 bottle	1 bottle	+4°C
Neutralization Buffer	1 bottle	1 bottle	+4°C
Collection Tubes (15mL)	4 tubes	12 tubes	+4°C

Reagents are ready to use as supplied.

# 3. Technical Considerations

#### 3.1 Amount of antibody that can be purified:

The antibody to be purified or cleaned up ideally is in a volume of 100 to 500  $\mu L.$  20 to 500  $\mu g$  of antibody can be purified in each run.

#### 3.2 Compatibility with Nanoparticle Conjugation Kits

Antibodies purified using the Antibody Purification Kit – Nanoparticles are fully compatible with our Gold, Latex, Europium and Magnetic particle conjugation kits, providing the antibody is purified and resuspended at a sufficient concentration for the conjugation reaction. We recommend a stock concentration of purified antibody of 1 mg/mL. For more information on required concentrations, consult the protocol for the applicable conjugation kit.

#### 3.3 Protein A affinity for immunoglobulins

Species	lgG	Binding strength
Rabbit	lgG	High
Human	lgG	High
Pig	lgG	High
Mouse	lgG <sub>2a</sub>	High
Mouse	lgG <sub>2b</sub>	High
Mouse	lgG1	Low/ Medium
Mouse	lgG <sub>3</sub>	Low/ Medium
Goat	lgG	Low
Sheep	lgG	Low
Rat	lgG	Low

#### 3.4 Test for protein:

Wherever possible, protein values should be determined using an absorbance at 280 nm. An extinction co-efficient of 1.4 is generally used for IgG – so a 1 mg/mL solution of IgG will give an absorbance value of 1.4 when measured with a 1cm path length.

When other methods are used such as BCA or Bradford protein assays, determinations should be performed before the addition of the Neutralization Buffer, as this can interfere with these reagents. Remove an aliquot for protein determination and neutralize the rest of the fraction immediately as the low pH of the Elution Buffer can denature the antibody.

When using Bradford-type reagents it is important to use an IgG standard curve. Failure to do this will result in incorrect antibody levels being calculated. If IgG is not available then a BSA standard curve can be used, but the IgG levels will be under-estimated by a factor of 2.3.

## 4. Assay Procedure

#### 4.1 Reconstitution of Protein A resin:

Add 300  $\mu$ L of Wash Buffer to each vial of Protein A Resin, mix by inversion for a few seconds and transfer to the spin cartridge. Spin for 30 seconds in a microfuge.

#### 4.2 Incubating sample with the resin:

To the antibody, add an appropriate amount of 10X Binding Buffer. For example, if the sample volume is 200  $\mu$ L, add 20  $\mu$ L of Binding Buffer. Pipette the sample into the Spin Cartridge and cap the tube. Incubate for 3 hours with agitation (end-over-end mixing or periodic shaking) at room temperature.

**\Delta Note:** The volume of antibody to be purified or cleaned up should ideally be 100-500 µL, though larger volumes may be processed by first incubating the antibody sample with the Protein A Resin in a larger vessel (e.g. 2 mL tube) prior to transferring to the Spin Cartridge.

#### 4.3 Wash procedure:

Microfuge the Spin Cartridge assembly for 30 seconds to remove most of the non-bound protein. Add 500  $\mu$ L of Wash Buffer and spin again. Repeat the wash procedure three times.

△ Note: Save the non-bound and wash fractions by transferring the material from the collecting tube after each spin to a set of microfuge tubes (not supplied). Do not use the four/twelve collecting tubes supplied with the kit, as these have an extended hinge to accommodate the spin cartridge, and are required for the elution step

#### 4.4 Elution:

Please see the Technical Consideration section on Test for protein before starting this step.

Transfer the cartridge to a clean collecting tube. Add 100  $\mu$ L of elution buffer and incubate for 2 minutes at room temperature with gentle agitation. Microfuge for 30 seconds.

Remove the collecting tube and add 25  $\mu$ L Neutralization Buffer to the tube. The Neutralization Buffer should be added to the sample as soon as possible, as long exposure to the low pH of the Elution Buffer can denature the antibody.

Place the cartridge in a new collecting tube and add a further 100  $\mu$ L of Elution buffer to the Protein A Resin. Incubate for 2 minutes at room temperature with gentle agitation. Spin and collect and neutralize as before.

Repeat the elution procedure until all four clean collecting cups have been used.

Pool the tubes with most protein (normally two tubes; if more than two tubes are strongly positive it is possible that you have used too much sample in your protein assay). However, if your application does not require a high concentration of antibody you may choose to pool all tubes that contain protein, regardless of concentration.

△ Note: The eluted antibody must be neutralized as soon as possible to avoid prolonged exposure to low pH of Elution Buffer which can result in denaturation of the IgG.

△ Note: The protein normally elutes in tubes 1 and 2, by spinpurification, and in tubes 2 and 3 by gravity-purification, but you should confirm this using a test for protein before pooling any of the tubes.

#### 4.5 Antibody storage:

Store at 4°C. Other storage conditions (e.g. frozen at -70°C) may also be satisfactory. The sensitivity of any antibody to freeze thaw should be determined by experimentation on small aliquots.

# **Technical Support**

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#### Austria

wissenschaftlicherdienst@abcam.com | 019-288-259 France supportscientifique@abcam.com | 01.46.94.62.96 Germany wissenschaftlicherdienst@abcam.com | 030-896-779-154 Spain soportecientifico@abcam.com | 91-114-65-60

#### Switzerland

technical@abcam.com Deutsch: 043-501-64-24 | Français: 061-500-05-30 **UK, EU and ROW** technical@abcam.com | +44(0)1223-696000

#### Canada

ca.technical@abcam.com | 877-749-8807 US and Latin America us.technical@abcam.com | 888-772-2226

#### Asia Pacific

hk.technical@abcam.com | (852) 2603-6823 China cn.technical@abcam.com | 400 921 0189 | +86 21 2070 0500

#### Japan

technical@abcam.co.jp | +81-(0)3-6231-0940 Singapore sg.technical@abcam.com | 800 188-5244

#### Australia

au.technical@abcam.com | +61-(0)3-8652-1450 New Zealand nz.technical@abc.com | +64-(0)9-909-7829