

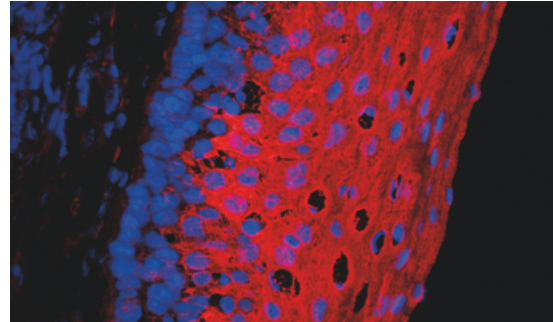
## VectaFluor™ Excel Amplified Kit

### Anti-Mouse IgG, DyLight® 594

**Cat. No.:** DK-2594

**Storage:** 2-8 °C

Instructions for immunofluorescent staining.



Tonsil: Multi-Cytokeratin detected with VectaFluor Excel Amplified Kit, Anti-Mouse IgG, DyLight 594 (red). Mounted in VECTASHIELD® HardSet™ Antifade Mounting Medium with DAPI (blue).

### DESCRIPTION

VectaFluor Excel Amplified Fluorescent Staining System offers a non-biotin amplification method for fluorescence applications. This system uses a proprietary, ready-to-use (R.T.U.) amplifier antibody, followed by a R.T.U. VectaFluor DyLight dye-labeled detection system.

### KIT COMPONENTS

Product Name	Volume
Normal Horse Serum, 2.5%	15 ml
Amplifier Antibody (Goat Anti-Mouse IgG)	15 ml
VectaFluor DyLight 594 Horse Anti-Goat IgG	15 ml

The VectaFluor Excel Amplified Kit will stain approximately 150 sections based on 100 µl per section.

### STORAGE:

- Store reagents in original bottles at 2-8 °C
- Do not freeze.

### PREPARATION OF WORKING SOLUTIONS

VectaFluor Excel Amplified Kit reagents are ready-to-use — no mixing or titering is necessary to obtain optimal staining.

The staining procedure should be performed at room temperature (20-25°C). VectaFluor Excel Amplified Kit reagents should be equilibrated to room temperature for optimal performance.

A number of different wash buffers can be used. One of the most common is 10 mM sodium phosphate, pH 7.5, 0.9% saline (PBS). 0.1% Tween 20 detergent may be added to the wash buffer and is especially recommended for use with automated stainers.

### STAINING PROCEDURE

1. For paraffin sections, deparaffinize and hydrate tissue sections through xylenes or other clearing agents and graded alcohol series.

For frozen sections or cell preparations fix with acetone or an appropriate fixative for the antigen under study, if required.

2. If antigen unmasking is required, perform this procedure using a Vector® Antigen Unmasking Solution, Citrate-based (H-3300) or Tris-based (H-3301).
3. Wash in buffer for 5 minutes.
4. Incubate for 20 minutes with 2.5% Normal Horse Serum.
5. Tip off excess serum from sections.
6. Incubate with mouse primary antibody diluted in an appropriate diluent.
7. Wash in buffer for 5 minutes.
8. Incubate for 15 minutes with Amplifier Antibody.
9. Wash in buffer for 5 minutes .
10. Incubate for 30 minutes with VectaFluor Reagent.
11. Wash for 2 x 5 minutes in buffer.
12. Mount in a media suitable for fluorescence, such as one of the VECTASHIELD Antifade Mounting Media.

Detailed product listing, specifications, protocols and additional information are available on our website: [vectorlabs.com](http://vectorlabs.com)