



Toxoplasma gondii IgM ELISA Kit

Catalog Number KA0226

96 assays

Version: 02

Intended for research use only

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Introduction

Intended Use

The *Toxoplasma gondii* IgM ELISA Kit is intended for use in the detection of IgM to *Toxoplasma gondii* in human serum.

Background

Toxoplasmosis is caused by the intracellular parasite *Toxoplasma gondii* and may be contracted by consuming contaminated meat or by coming in contact with cat feces containing oocysts. In adolescence and adulthood, most infections are subclinical. However, if a pregnant woman contracts toxoplasmosis, it may be passed through the placenta to the fetus, resulting in congenital toxoplasmosis, which is a cause of mortality and malformation. Asymptomatic infants may develop abnormalities later in life. The *Toxoplasma gondii* IgM ELISA Kit is an accurate serologic method to detect *Toxoplasma* IgM antibody for identification of toxoplasmosis.

Principle of the Assay

Purified *Toxoplasma* antigen is coated on the surface of microwells. Diluted serum is added to the wells, and the *Toxoplasma* IgM-specific antibody, if present, binds to the antigen. All unbound materials are washed away. HRP-conjugate is added, which binds to the antibody-antigen complex. Excess HRP-conjugate is washed off and a solution of TMB Reagent is added. The enzyme conjugate catalytic reaction is stopped at a specific time. The intensity of the color generated is proportional to the amount of IgM-specific antibody in the sample. The results are read by a microwell reader compared in a parallel manner with calibrator and controls.

General Information

Materials Supplied

List of component

Component	Amount
Microtiter Wells: Purified <i>Toxoplasma</i> antigen-coated wells	96 (8 x 12) wells
Enzyme Conjugate Reagent (red color); Red cap	12 mL
Sample Diluent (blue color)	22 mL
Negative Control: Range stated on label. Natural cap.	100 µL/vial
Cut-off Calibrator: Yellow cap. <i>Toxoplasma M</i> , Index = 1.	100 µL/vial
Positive Control: Range stated on label. Red cap.	100 µL/vial
Wash Buffer Concentrate (20x)	50 mL
TMB Reagent (One-Step)	11 mL
Stop Solution (1N HCl). Natural cap.	11 mL

Storage Instruction

- ✓ Store the kit at 2-8°C.
- ✓ Keep microwells sealed in a dry bag with desiccants.
- ✓ The reagents are stable until expiration of the kit.
- ✓ Do not expose test reagents to heat, sun or strong light during storage or usage.

Precautions for Use

- ✓ Warning and precautions
 - Potential biohazardous materials: The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, as there is no test method that can offer complete assurance that HIV, hepatitis B virus or other infectious agents are absent, these reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories." 1984.
 - Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
 - The components from different lots should not be mixed.
 - This product contains components preserved with sodium azide. Sodium azide may react with lead and copper plumbing to form explosive metal azide. On disposal, flush with a large volume of water.

✓ Limitations of the procedures

- Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the package insert instructions and with adherence to good laboratory practice.
- The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
- Serum samples demonstrating gross lipemia, gross hemolysis, or turbidity should not be used with this test.

Assay Protocol

Reagent Preparation

- ✓ All reagents should be allowed to reach room temperature (18-25°C) before use.
- ✓ Dilute 1 volume of Wash Buffer (20x) with 19 volumes of distilled water. For example, dilute 50 mL of Wash Buffer (20x) into distilled water to prepare 1000 mL of Wash Buffer (1x). Wash Buffer is stable for 1 month at 2-8°C. Mix well before use.

Sample Preparation

- ✓ Collect blood specimens and separate the serum.
- ✓ Specimens may be refrigerated at 2-8°C for up to 7 days or frozen for up to 6 months. Avoid repetitive freezing and thawing of serum sample.

Assay Procedure

1. Place the desired number of coated wells into the holder.
2. Prepare 1:40 dilution of test samples, Negative Control, Positive Control, and Calibrator by adding 5 µL of the sample to 200 µL of Sample Diluent. Mix well.
3. Dispense 100 µL of diluted sera, Calibrator, and Controls into the appropriate wells. For the reagent blank, dispense 100 µL Sample Diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well.
4. Incubate at 37°C for 30 minutes.
5. At the end of incubation period, remove liquid from all wells. Rinse and flick the microtiter wells 5 times with diluted Wash Buffer (1x).
6. Dispense 100 µL of Enzyme Conjugate to each well. Mix gently for 10 seconds.
7. Incubate at 37°C for 30 minutes.
8. Remove Enzyme Conjugate from all wells. Rinse and flick the microtiter wells 5 times with diluted Wash Buffer (1x).
9. Dispense 100 µL of TMB Reagent into each well. Mix gently for 10 seconds.
10. Incubate at 37°C for 15 minutes.
11. Add 100 µL of Stop Solution (1N HCl) to stop reaction.
12. Mix gently for 30 seconds. It is important to make sure that all the blue color changes to yellow color completely. *Note: Make sure there are no air bubbles in each well before reading.*
13. Read O.D. at 450 nm within 15 minutes with a microwell reader.

Data Analysis

Calculation of Results

1. Calculate the mean of duplicate cut-off calibrator value X_c .
2. Calculate the mean of duplicate positive control (X_p), negative control (X_n) and samples (X_s).
3. Calculate the Toxoplasma IgM Index of each determination by dividing the mean values of each sample (X) by calibrator mean value, X_c .

✓ Example of typical results:

Note: The O.D. values are for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data.

Cut-off Calibrator Toxoplasma M Index = 1.0

1. Cut-off Calibrator O.D. = 0.952, 0.885 $X_c = 0.919$
2. Negative Control O.D. = 0.066, 0.065 $X_n = 0.066$
Toxo M Index = $X_n / X_c = 0.066 / 0.919 = 0.07$
3. Positive Control O.D. = 1.267, 1.276 $X_p = 1.272$
Toxo M Index = $X_p / X_c = 1.272 / 0.919 = 1.38$
4. Patient sample O.D. = 1.679, 1.665 $X_s = 1.672$
Toxo M Index = $X_s / X_c = 1.672 / 0.919 = 1.82$

✓ Quality control

The test run may be considered valid provided the following criteria are met:

1. The O.D. value of the reagent blank against air from a microwell reader should be less than 0.250.
2. If the O.D. value of the Cut-off Calibrator is lower than 0.250, the test is not valid and must be repeated.
3. The Toxo M Index for Negative and Positive Controls should be in the range stated on the Certificate of Analysis.

✓ Interpretation

- Negative: Toxo M Index less than 0.90 is negative for IgM antibody to *T. gondii*.
- Equivocal: Toxo M Index between 0.91-0.99 is equivocal. Sample should be retested.
- Positive: Toxo M Index of 1.00 or greater is positive for IgM antibody to *T. gondii* and indicates the probability of current or recent toxoplasmosis.

Performance Characteristics

- ✓ Specificity and Sensitivity:
- A total of 159 samples were used to evaluate specificity and sensitivity of the test. The Abnova's Toxo IgM ELISA test results were compared to a commercial ELISA kit results:

		Reference Toxo IgM ELISA			
		N	E	P	Total
Abnova Toxo IgM ELISA	N	130(D)	2	0(B)	132
	E	1	1	0	2
	P	4(C)	2	19(A)	25
	Total	135	5	19	159

$$\text{Sensitivity} = A / (A+B) = 19 / 19 = 100.0\%$$

$$\text{Specificity} = D / (C+D) = 130 / 134 = 97.0\%$$

$$\text{Accuracy} = (A+D) / (A+B+C+D) = 149 / 153 = 97.4\%$$

- ✓ Precision:
- Intra-Assay Precision

Within run precision was determined by replicate determinations of three different serum samples in one assay. Within-assay variability is shown below:

Sample	1	2	3
# Reps.	24	24	24
Mean A_{450}	2.628	1.103	0.090
S.D. (A_{450})	0.052	0.022	0.002
C.V. (%)	2.0	2.0	1.9

- Inter-Assay Precision

Between-run precision was determined by replicate measurements of three different serum samples over a series of individually calibrated assays. Between-assay variability is shown below:

Sample	1	2	3
# Reps.	20	20	20
Mean A_{450}	2.534	0.988	0.101
S.D. (A_{450})	0.060	0.026	0.003
C.V. (%)	2.4	2.7	3.2

Resources

References

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2. Lin, T.M., S.P. Halbert and G.R. O'Connor. Standardized Quantitative Enzyme-linked Immunoassay for Antibodies to Toxoplasma Gondii. J. Clin. Microbiol. Vol. 11, 6:675-681, 1980.
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4. Voller, A., D.E. Bidwell, A. Bartlett, D.G. Flick, M. Perkins and B. Oladshin. A Microplate Enzyme-immunoassay for Toxoplasma Antibodies. J. Clin. Path. 29:150-153, 1976.

Plate Layout

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1								
	A	B	C	D	E	F	G	H