cGMP Direct Immunoassay Kit

Catalog Number KA0887

100 assays

Version: 04

Intended for research use only

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Table of Contents

Introduction **3**

Background 3

General Information **4**

Materials Supplied 4

Storage Instruction 4

Precautions for Use 5

Assay Protocol **6**

Reagent Preparation 6

Sample Preparation 6

Assay Procedure 6

Data Analysis **8**

Calculation of Results 8

Introduction

Background

Adenosine and guanosine 3',5'-cyclic monophosphate (cAMP and cGMP) are important "second messengers" involved in many physiological processes. The cGMP Direct Immunoassay Kit provides a direct competitive immunoassay for sensitive and quantitative determination of cGMP level in biological samples. The kit utilizes the recombinant Protein G coated plate to anchor cGMP polyclonal antibody. cGMP-HRP conjugates directly competes with cGMP from samples for binding to the cGMP specific antibody on the plate. After incubation and washing, the amount of cGMP-HRP bound to plate can easily be determined by reading $OD_{450\ nm}$. The intensity of $OD_{450\ nm}$ is inversely proportional to the concentration of cGMP in samples. The kit provides a new acetylation procedure that improves detection signal significantly. The kit can detect 0.04 -10 pmol/well (0.008 - 2 µM) cGMP samples.

General Information

Materials Supplied

List of component

Component	Amount
10X cGMP Assay Buffer	25 mL
Standard cGMP (10 nmol)	1 vial
Neutralizing Buffer	7.5 mL
Acetylating Reagent A	0.75 mL
Acetylating Reagent B	1.5 mL
Anti-cGMP pAb/BSA	1 vial
cGMP-HRP/BSA	1 vial
HRP Developer	10 mL
Protein G Coated Plate	1 each

Storage Instruction

The kit should be stored at -20°C. After reconstitution, some components may be stored at 4°C as instructed above, stable for up to 1- 2 months.

Component	Storage
10X cGMP Assay Buffer	4°C
Standard cGMP (10 nmol)	-20°C
Neutralizing Buffer	4°C
Acetylating Reagent A	4°C
Acetylating Reagent B*	4°C
Anti-cGMP pAb/BSA	-20°C
cGMP-HRP/BSA	-20°C
HRP Developer	4°C
Protein G Coated Plate	-20°C

**Note: Acetylating Reagent B is very volatile and hence the vial has to be tightly capped and stored only at 4°C.*

Precautions for Use

- ✓ For research use only! Not to be used on humans.
- ✓ General Consideration
- cGMP samples in 0.1 M HCl (final concentration) is stable and can be used directly in the assay. Make dilutions of your sample with 0.1 M HCl to the range of 0.04-10 pmol/well (0.008-2 µM).
- Plasma, serum, whole blood, and tissue homogenates often contain phosphodiesterases and large amount of immunoglobulins (Igs) which may interfere with the assay. However, preparing samples in 0.1M HCl can generally inactivate phosphodiesterases and lower the concentration of Igs, making the samples suitable for the assay. Both phosphodiesterases and Igs can also be removed by 5% TCA precipitation or by using 10 Kd molecular weight cut off microcentrifuge filters.
- To determine whether interference is presence in your sample, you may make two different dilutions. If the two different dilutions of sample show good correlation in the final calculated cGMP concentrations, purification is not required. If you do not see good correlation of the different dilutions, deproteinize the sample by using TCA or 10 Kd molecular cut off microcentrifuge filters.
- Some organic solvents may interfere with the assay and may need to be removed prior to the assay.

Assay Protocol

Reagent Preparation

- ✓ Dilute the 10X cGMP Assay Buffer to 1X Assay Buffer with MilliQ water. Store at 4°C.
- ✓ Reconstitute the Standard cGMP (pellet may not be visible) in 1 mL of 0.1 M HCl (not provided), vertex for 10 seconds to generate 10 pmol/µL cGMP stock standard solution.
- ✓ Reconstitute rabbit anti-cGMP pAb and cGMP-HRP each with 1.1 mL of the 1X Assay Buffer as stock solutions.
- ✓ Unused well strips can be kept at -20°C with the desiccants, stable for up to 1 month.
- ✓ The kit should be stored at -20°C. After reconstitution, some components may be stored at 4°C as instructed above, stable for up to 1- 2 months.
- ✓ **NOTE- Acetylating Reagent B is very volatile and hence the vial has to be tightly capped and stored only at 4°C.*

Sample Preparation

- ✓ Urine, Plasma and Culture Media: Urine, plasma, and culture media may be tested directly after adding 1/10 volume of 1M HCl, and remove precipitates if occur.
- ✓ Culture Cells: For suspension cells collect by centrifugation. Add 1 mL of 0.1 M HCl for every 35 cm^2 of surface area. (e.g., 10 cm plate at 70 % confluency is ~ 110 cm^2, so use ~ 3.1 mL). Incubate at room temperature for 20 minutes on ice. For adherent cells add the HCL directly, scrape cells off the surface. Dissociate sample by pipetting up and down until suspension is homogeneous. Transfer to a centrifuge tube and centrifuge at top speed for 10 min. The supernatant can be assayed directly. Protein concentration ≥ 1 mg/mL is recommended for reproducible results.
- ✓ Tissue Samples: Cyclic nucleotides may be metabolized quickly in tissue, so it is important to rapidly freeze tissues after collection (e.g., using liquid nitrogen). Weigh the frozen tissue and add 5-10 volume of 0.1 M HCl. Homogenize the sample on ice using a Polytron-type homogenizer. Spin at top speed for 5 min and collect the supernatant. The supernatant may be assayed directly.

Assay Procedure

- ✓ Prepare cGMP Standard Curve and Samples:

1. Add 200 µL of the 10 pmol/µL standard cGMP stock into 800 µL of 0.1 M HCl to generate 2 pmol/µL cGMP working solution. The diluted cGMP should be used within 1 hour.
2. Label 11 microcentrifuge tubes, 10, 5, 2.5, 1.25, 0.625, 0.3125, 0.156, 0.078, 0.039, 0 pmol/50 µL. *(Note: these concentrations represent what will finally be in the wells after the dilutions mentioned below).*
3. Add 200 µL of the 2 pmol/µL cGMP into the tube labeled 10 pmol (enough for 20 tests), add100 µL 0.1M HCl into the rest of tubes.

4. Transfer 100 µL from the 10 pmol tube into the labeled 5 pmol tube, mix. Continue the serial dilution by transfer 100 µL from the 5 pmol tube to 2.5, 1.25, 0.625, 0.3125, 0.156, 0.078, 0.039 pmol tubes. Discard the 100 µL mixture from the last 0.039 pmol tube. The diluted cGMP should be used within 1 hour.
5. Label new tubes for test samples, add 100 µL each test sample per tube. We suggest using different dilutions for each sample (dilute with 0.1 M HCl).
6. Add 50 µL of Neutralizing Buffer to each tube to neutralize the HCl in the samples and standards.
7. Prepare Acetylating Reagent Mix *(Note: 5 µL is needed for each assay)*: Mix 1 volume of Acetylating Reagent A with two volumes of Acetylating Reagent B in a microtube. Prepare just enough for the experiment. Use within 1 hour.
8. Add 5 µL of the Acetylating Reagent Mix directly into each test solution (both standard and sample), IMMEDIATELY vertex 2-3 seconds following each addition without delay, one tube at a time and incubate at room temperature for 10 min.
9. Add 845 µL 1X Assay Buffer into each tube, mix well. Use for below quantification.

Note: The acetylation step improves the assy sensitivity significantly and avoid the interferences of many components in unpurified samples. (If cGMP in your samples are very low, the acetylation reagents can be dried after step 8, without dilution step 9 to minimize the volume. Then reconstituted in a 50 -100 µL volume of Assay Buffer).

✓ Quantification cGMP

1. Add 50 µL of the acetylated Standard cGMP and test samples from Step 9 above to each well of the Protein G coated 96-well plate. We suggest duplicate assays for each sample and standard.
2. Add 10 µL of the reconstituted cGMP antibody per well to the standard cGMP and sample wells except the well with 0 pmol cGMP. *(Note: Do not add cGMP antibody into the well with 0 pmol cGMP, instead add 10 µL of 1X Assay Buffer for background reading).* Incubate for 1 hour at room temperature with gentle agitation.

   Note: Using a repeating pipette is recommended for minimizing pippetting errors.
3. Add 10 µL of cGMP-HRP to each well and incubate for 1 hr at room temperature with gentle agitation.
4. Wash 5 times with 200 µL 1X Assay Buffer each time. Completely empty the wells by tapping the plate on a fresh paper towel after each wash step.
5. Add 100 µL of HRP developer and develop for 1 hour at room temperature with agitation.
6. Stop the reaction by adding 100 µL of 1 M HCl (not provided) to each well (sample color should change from blue to yellow).
7. Read sample at OD 450 nm.
8. Subtract OD450 nm background reading (the well with 0 pmol cGMP) from all samples and standards. Plot standard curve to observe the linear portion, then replot only the linear portion and in Excel add a trendline, then use the trend line linear formula (y=mx+b). Calculate amount of cGMP in samples after correcting the for dilution factors.

Data Analysis

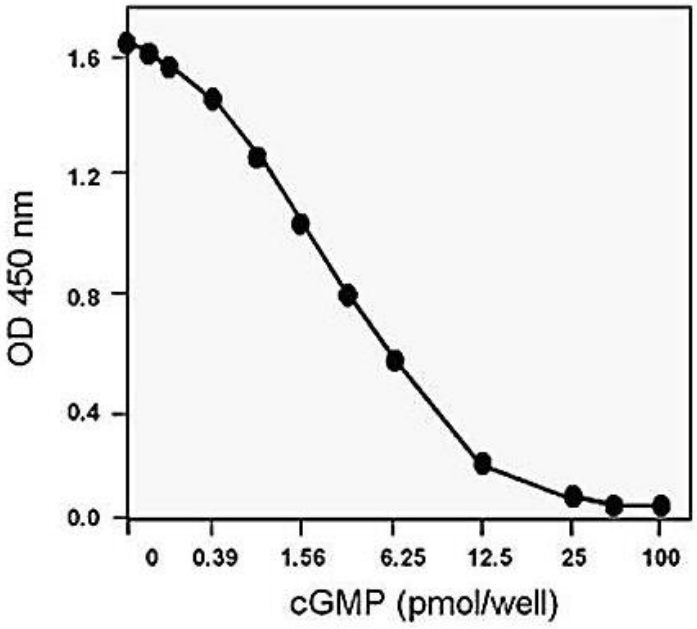
Calculation of Results

Calculations:

C = Sa/Sv pmol/µl or nmol/ml or µM.

Where: Sa is the cGMP amount (pmol) from the Standard Curve.

Sv is the sample volume (µL) added into the assay wells after dilution factor correction.



cGMP Standard Curve: The assay was performed following the kit protocol.