Thyroglobulin Ab ELISA Kit

Catalog Number KA0951
96 assays
Version: 04

Intended for research use only
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Introduction

Intended Use

The Thyroglobulin Ab ELISA Kit is intended for the detection of IgG antibody to Thyroglobulin (TG) in human serum or plasma.

Background

Thyroglobulin is a water soluble glycoprotein that is involved in the storage and synthesis of thyroid hormones. The thyroid microsomal antigen has been shown to be the enzyme thyroid peroxidase (TPO). Antibodies to thyroglobulin and or microsomal antigen are present in most patients with goitrous thyroiditis (Hashimoto disease), atrophic thyroiditis and about 70-90% of Graves disease. Antibodies are also found in about half of the patients with primary hypothyroidism and thyrotoxicosis, and 10-20% of patients with simple goiters and thyroid tumors. There is also a relationship between thyroid antibodies and diabetes mellitus. Thyroid autoantibodies are present in about 6-7% of normals and their incidence increases with age. Classically, autoantibodies to thyroid antigens are detected by precipitation reactions, hemagglutination and by immunofluorescence. However the tests are subjective and lack high sensitivity. Enzyme-Linked Immunosorbent Assays (ELISAs) combine greater sensitivity, objective reading and ease of use. ELISAs have been developed and validated for detecting autoantibodies to thyroid antigens.

Principle of the Assay

Diluted serum samples are added to wells coated with purified TG recombinant antigen. TG specific antibody, if present, binds to the antigen. All unbound materials are washed away and the enzyme conjugate is added to bind to the antibody-antigen complex, if present. Excess enzyme conjugate is washed off and substrate is added. The plate is incubated to allow the hydrolysis of the substrate by the enzyme. The intensity of the color generated is proportional to the amount of TG specific antibody in the sample.
General Information

Materials Supplied

List of component

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microwells coated with TG antigen</td>
<td>96 (8x12) wells</td>
</tr>
<tr>
<td>Sample Diluent (ready to use)</td>
<td>22 mL</td>
</tr>
<tr>
<td>Calibrator (ready to use)</td>
<td>1 mL</td>
</tr>
<tr>
<td>Positive Control (ready to use)</td>
<td>1 mL</td>
</tr>
<tr>
<td>Negative Control (ready to use)</td>
<td>1 mL</td>
</tr>
<tr>
<td>Enzyme conjugate (ready to use)</td>
<td>12 mL</td>
</tr>
<tr>
<td>TMB Substrate (ready to use)</td>
<td>12 mL</td>
</tr>
<tr>
<td>Stop Solution (ready to use)</td>
<td>12 mL</td>
</tr>
<tr>
<td>Wash concentrate 20X</td>
<td>25 mL</td>
</tr>
</tbody>
</table>

Storage Instruction

✓ Store the kit at 2-8°C.
✓ Keep microwells sealed in a dry bag with desiccants.
✓ The reagents are stable until expiration of the kit.
✓ Do not expose test reagents to heat, sun or strong light.

Materials Required but Not Supplied

✓ Distilled or deionized water
✓ Precision pipettes
✓ Disposable pipette tips
✓ ELISA reader capable of reading absorbance at 450 nm
✓ Absorbance paper or paper towel
✓ Graph paper

Precautions for Use

• Limitations of the procedure
✓ Lipemic or hemolyzed samples may cause erroneous results.
• Warning and Precautions
✓ Potential biohazardous materials:
  The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent. These reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, “Biosafety in Microbiological and Biomedical Laboratories” 1984.
✓ This kit is designed for research use only.
✓ Optimal results will be obtained by strict adherence to the test protocol. Precise pipetting as well as following the exact time and temperature requirements is essential.
✓ Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
✓ The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
✓ This product contains components preserved with sodium azide. Sodium azide may react with lead and copper plumbing to form explosive metal azide. On disposal, flush with a large volume of water.
Assay Protocol

Reagent Preparation

Prepare 1X Wash buffer by adding the contents of the bottle (25 mL, 20X) to 475 mL of distilled or deionized water. Store at room temperature (20-25°C).

Sample Preparation

✓ Collect blood specimens and separate the serum.
✓ Specimens may be refrigerated at 2–8 °C for up to seven days or frozen for up to six months. Avoid repetitive freezing and thawing.

Assay Procedure

Bring all specimens and kit reagents to room temperature (20-25°C) and gently mix.

1. Place the desired number of coated strips into the holder.
2. Negative control, positive control, and calibrator are ready to use. Prepare 1:21 dilution of test samples, by adding 10 μL of the sample to 200 μL of sample diluent. Mix well.
3. Dispense 100 μL of diluted sera, calibrator and controls into the appropriate wells. For the reagent blank, dispense 100 μL sample diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well. Incubate for 20 minutes at room temperature.
4. Remove liquid from all wells. Wash wells three times with 300 μL of 1X wash buffer. Blot on absorbance paper or paper towel.
5.Dispense 100 μL of enzyme conjugate to each well and incubate for 20 minutes at room temperature.
6. Remove enzyme conjugate from all wells. Wash wells three times with 300 μL of 1X wash buffer. Blot on absorbance paper or paper towel.
7. Dispense 100 μL of TMB substrate and incubate for 10 minutes at room temperature.
8. Add 100 μL of stop solution.
9. Read O.D. at 450 nm using ELISA reader within 15 min. A dual wavelength is recommended with reference filter of 600-650 nm.
Data Analysis

Calculation of Results

✓ Check Calibrator Factor (CF) value on the calibrator bottle. This value might vary from lot to lot. Make sure you check the value on every kit.
✓ Calculate the cut-off value: Calibrator OD x Calibrator Factor (CF).
✓ Calculate the Ab (Antibody) Index of each determination by dividing the O.D. value of each sample by cut-off value

- Example of typical results:
  Calibrator mean OD = 0.8
  Calibrator Factor (CF) = 0.5
  Cut-off Value = 0.8 x 0.5 = 0.400
  Positive control O.D. = 1.2
  Ab Index = 1.2 / 0.4 = 3
  Test sample O.D. = 1.6
  Ab Index = 1.6 / 0.4 = 4.0

- Quality Control
  The test run may be considered valid provided the following criteria are met:
  1. The O.D. of the Calibrator should be greater than 0.250.
  2. The Ab index for Negative control should be less than 0.9.
  3. The Ab Index for Positive control should fall within the range specified on COA/label.

✓ Interpretation
  The following is intended as a guide to interpretation of TG antibody test results; each laboratory is encouraged to establish its own criteria for test interpretation based on sample populations encountered.

- Antibody Index Interpretation
  <0.9  No detectable TG antibody by ELISA.
  0.9-1.1 Borderline positive. Follow-up testing is recommended.
  >1.1  detectable TG antibody by ELISA.

- Converting of Ab Index to IU/mL
  As an option, TG Ab index may be converted to IU/mL by multiplying Ab index value by 100. International units may then be interpreted as follows:
  <90 IU/mL:  Negative
  90-110 IU/mL: Borderline positive
  >110 IU/mL:  Positive
**Performance Characteristic**

- **Sensitivity and Specificity**
  121 donor sera were tested by this ELISA and a reference ELISA methods. 28 were positive and 88 were negative by both methods (96% agreement). The results are summarized below:

<table>
<thead>
<tr>
<th>TG Ab ELISA</th>
<th>+</th>
<th>-</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reference ELISA kit</td>
<td>28</td>
<td>3</td>
<td>31</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>88</td>
<td>90</td>
</tr>
<tr>
<td>Total</td>
<td>30</td>
<td>91</td>
<td>121</td>
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</table>

- **Precision**
  **Intra-Assay Study**

<table>
<thead>
<tr>
<th>Serum</th>
<th>No. of Replicates</th>
<th>Mean</th>
<th>Standard Deviation</th>
<th>Coefficient of Variation %</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>16</td>
<td>1.56</td>
<td>0.12</td>
<td>7.7</td>
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<tr>
<td>2</td>
<td>16</td>
<td>0.84</td>
<td>0.06</td>
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<tr>
<td>3</td>
<td>16</td>
<td>0.22</td>
<td>0.02</td>
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  **Inter-Assay Study**

<table>
<thead>
<tr>
<th>Serum</th>
<th>No. of Replicates</th>
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<th>Coefficient of Variation %</th>
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<tr>
<td>1</td>
<td>10</td>
<td>1.47</td>
<td>0.14</td>
<td>9.5</td>
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<td>2</td>
<td>10</td>
<td>0.97</td>
<td>0.09</td>
<td>9.3</td>
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<td>3</td>
<td>10</td>
<td>0.24</td>
<td>0.03</td>
<td>12.5</td>
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References


3. Franke WG; Schimming C; Wunderlich G. Can thyroid peroxidase be used as a complementary tumor marker besides thyroglobulin? Preliminary experience with determination of TPO in differentiated thyroid carcinomas. Anticancer Res 1997; 17(4B):2999-3002.


7. Nakamura H; Genma R; Mikami T; Kitahara A; Natsume H; Andoh S; Nagasawa S; Nishiyama K; Chida K; Sato A; Yoshimi T. High incidence of positive autoantibodies against thyroid peroxidase and thyroglobulin in patients with sarcoidosis. Clin Endocrinol (Oxf) 1997; 46(4):467-72.

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