



# KLK3 (Human) ELISA Kit

Catalog Number KA1063

96 assays

Version: 04

Intended for research use only

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## **Introduction**

### **Intended Use**

For quantitative detection of human KALLIKREIN 3 in serum, body fluids, tissue lysates or cell culture supernates.

### **Background**

Prostate-specific antigen (PSA), also known as kallikrein III, seminin, semenogelase,  $\gamma$ -seminoprotein and P-30 antigen) is a 34 kD glycoprotein manufactured almost exclusively by the prostate gland; PSA is produced for the ejaculate where it liquifies the semen in the seminal coagulum and allows sperm to swim freely.<sup>1</sup> It is also believed to be instrumental in dissolving the cervical mucous cap, allowing the entry of sperm.<sup>2</sup> It is a serine protease enzyme, the gene of which is located on the nineteenth chromosome (19q13).<sup>3</sup>

### **Principle of the Assay**

KLK3 (Human) ELISA Kit was based on standard sandwich enzyme-linked immune-sorbent assay technology. Human KALLIKREIN 3 specific-specific polyclonal antibodies were precoated onto 96-well plates. The human specific detection polyclonal antibodies were biotinylated. The test samples and biotinylated detection antibodies were added to the wells subsequently and then followed by washing with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex was added and unbound conjugates were washed away with PBS or TBS buffer. HRP substrate TMB was used to visualize HRP enzymatic reaction. TMB was catalyzed by HRP to produce a blue color product that changed into yellow after adding acidic stop solution. The density of yellow is proportional to the human KALLIKREIN 3 amount of sample captured in plate.

## General Information

### Materials Supplied

List of component

Component	Amount
Lyophilized recombinant human KALLIKREIN 3 standard.	20 ng × 2
One 96-well plate precoated with anti- human KALLIKREIN 3 antibody.	96 (8x12) wells
Sample diluent buffer	30 mL
Biotinylated anti- human KALLIKREIN 3 antibody: dilution 1:100.	130 µL
Antibody diluent buffer	12 mL
Avidin-Biotin-Peroxidase Complex (ABC): dilution 1:100.	130 µL
ABC diluent buffer	12 mL
TMB color developing agent	10 mL
TMB stop solution	10 mL

### Storage Instruction

Store at 4°C for frequent use, at -20°C for infrequent use. Avoid multiple freeze-thaw cycles.

### Materials Required but Not Supplied

- ✓ Microplate reader in standard size.
- ✓ Automated plate washer.
- ✓ Adjustable pipettes and pipette tips. Multichannel pipettes are recommended in the condition of large amount of samples in the detection.
- ✓ Clean tubes and Eppendorf tubes.
- ✓ Washing buffer (neutral PBS or TBS).
- Preparation of 0.01M TBS:  
Add 1.2 g Tris, 8.5 g NaCl; 450 µL of purified acetic acid or 700 µL of concentrated hydrochloric acid to 1000 mL H<sub>2</sub>O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1 L.
- Preparation of 0.01 M PBS:  
Add 8.5 g sodium chloride, 1.4 g Na<sub>2</sub>HPO<sub>4</sub> and 0.2 g NaH<sub>2</sub>PO<sub>4</sub> to 1000 mL distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1 L.

### **Precautions for Use**

- ✓ To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
- ✓ The TMB Color Developing agent is colorless and transparent before using, contact us freely if it is not the case.
- ✓ Before using the Kit, spin tubes and bring down all components to the bottom of tubes.
- ✓ Duplicate well assay is recommended for both standard and sample testing.
- ✓ Don't let 96-well plate dry, for dry plate will inactivate active components on plate.
- ✓ Don't reuse tips and tubes to avoid cross contamination.
- ✓ To avoid to use the reagents from different batches together.
- ✓ In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution will be pre-warmed in 37°C for 30 min before using.

## Assay Protocol

### Reagent Preparation

- Reconstitution of the human KALLIKREIN 3 standard: KALLIKREIN 3 standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of KALLIKREIN 3 standard (20ng per tube) are included in each kit. Use one tube for each experiment.
  - ✓ 20 ng/mL of human KALLIKREIN 3 standard solution: Add 1 mL sample diluent buffer into one tube, keep the tube at room temperature for 10 min and mix thoroughly.
  - ✓ 10 ng/mL→0.312 ng/mL of human KALLIKREIN 3 standard solutions: Label 6 Eppendorf tubes with 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL, respectively. Aliquot 0.3 mL of the sample diluent buffer into each tube. Add 0.3 mL of the above 20 ng/ml KALLIKREIN 3 standard solution into 1st tube and mix. Transfer 0.3 mL from 1st tube to 2nd tube and mix. Transfer 0.3 mL from 2nd tube to 3rd tube and mix, and so on.

*Note: The standard solutions are best used within 2 hours. The 20 ng/mL standard solution may be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.*

- Preparation of biotinylated anti-human KALLIKREIN 3 antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.
  - ✓ The total volume should be: 0.1 mL/well x (the number of wells). (Allowing 0.1-0.2 mL more than total volume)
  - ✓ Biotinylated anti-human KALLIKREIN 3 antibody should be diluted in 1:99 with the antibody diluent buffer and mixed thoroughly.
- Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.
  - ✓ The total volume should be: 0.1mL/well x (the number of wells). (Allowing 0.1-0.2 mL more than total volume)
  - ✓ Avidin-Biotin-Peroxidase Complex (ABC) should be diluted in 1:99 with the ABC dilution buffer and mixed thoroughly.

### Sample Preparation

- Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

- ✓ Cell culture supernate, tissue lysate or body fluids: Remove particulates by centrifugation, analyze immediately or aliquot and store at -20°C

- ✓ Serum: Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 min. Analyze the serum immediately or aliquot and store frozen at -20°C.
- Sample Dilution Guideline

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. The sample must be well mixed with the diluents buffer.

  - ✓ High target protein concentration (200-2000 ng/mL). The working dilution is 1:100. i.e. Add 1 µL sample into 99 µL sample diluent buffer.
  - ✓ Medium target protein concentration (20-200 ng/mL). The working dilution is 1:10. i.e. Add 10 µL sample into 90 µL sample diluent buffer.
  - ✓ Low target protein concentration (0.312-20 ng/mL). The working dilution is 1:2. i.e. Add 50 µL sample to 50 µL sample diluent buffer.
  - ✓ Very Low target protein concentration ( $\leq 0.312$  ng/mL). No dilution necessary, or the working dilution is 1:2.

### **Assay Procedure**

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 min before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard KALLIKREIN 3 detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of KALLIKREIN 3 amount in samples.

1. Aliquot 0.1 mL per well of the 20 ng/mL, 10 ng/mL, 5 ng/mL, 2.5 ng/mL, 1.25 ng/mL, 0.625 ng/mL, 0.312 ng/mL human KALLIKREIN 3 standard solutions into the precoated 96-well plate. Add 0.1 mL of the sample diluent buffer into the control well (Zero well). Add 0.1 mL of each properly diluted sample of human serum, body fluids, tissue lysates or cell culture supernatants to each empty well. See "Sample Dilution Guideline" above for details. It is recommended that each human KALLIKREIN 3 standard solution and each sample be measured in duplicate.
2. Seal the plate with the cover and incubate at 37°C for 90 min.
3. Remove the cover, discard plate content, and blot the plate onto paper towels or other absorbent material. Do NOT let the wells completely dry at any time.
4. Add 0.1 mL of biotinylated anti-human KALLIKREIN 3 antibody working solution into each well and incubate the plate at 37°C for 60 min.
5. Wash plate 3 times with 0.01 M TBS or 0.01 M PBS, and each time let washing buffer stay in the wells for 1 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.  
(Plate Washing Method: Discard the solution in the plate without touching the side walls. Blot the plat

onto paper towels or other absorbent material. Soak each well with at least 0.3 mL PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of THREE washes. *Note: For automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the plate onto paper towels or other absorbent material.*)

6. Add 0.1mL of prepared ABC working solution into each well and incubate the plate at 37°C for 30 min.
7. Wash plate 5 times with 0.01 M TBS or 0.01 M PBS, and each time let washing buffer stay in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material. (See Step 5 for plate washing method).
8. Add 90 µL of prepared TMB color developing agent into each well and incubate plate at 37°C in dark for 20-25 min (*Note: For reference only, the optimal incubation time should be determined by end user. And the shades of blue can be seen in the wells with the four most concentrated human KALLIKREIN 3 standard solutions; the other wells show no obvious color*).
9. Add 0.1 mL of prepared TMB stop solution into each well. The color changes into yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 min after adding the stop solution.

- Summary

1. Add samples and standards and incubate the plate at 37°C for 90 min. Do not wash.
2. Add biotinylated antibodies and incubate the plate at 37°C for 60 min. Wash plate 3 times with 0.01 M TBS.
3. Add ABC working solution and incubate the plate at 37°C for 30 min. Wash plate 5 times with 0.01 M TBS.
4. Add TMB color developing agent and incubate the plate at 37°C in dark for 20-25 min.
5. Add TMB stop solution and read.



## Data Analysis

### Calculation of Results

For calculation, (the relative O.D.<sub>450</sub>) = (the O.D.<sub>450</sub> of each well) – (the O.D.<sub>450</sub> of Zero well). The standard curve can be plotted as the relative O.D.<sub>450</sub> of each standard solution (Y) vs. the respective concentration of the standard solution (X). The human KALLIKREIN 3 concentration of the samples can be interpolated from the standard curve.

*Note: if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.*

- Typical result

Typical Data Obtained from human KALLIKREIN 3

(TMB reagent incubate at 37°C for 20 min)

Concentration (ng/mL)	0.0	0.312	0.625	1.25	2.5	5	10	20
O.D	0.005	0.118	0.224	0.378	0.702	1.346	1.694	2.178

### Performance Characteristics

- Range  
0.312 ng/mL-20 ng/mL
- Sensitivity  
< 10 pg/mL
- Specificity  
No detectable cross-reactivity with any other cytokine.

## Resources

### References

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2. edited by Wayne J.G. Hellstrom. (1999). "Chapter 8: What is the prostate and what is its function?". American Society of Andrology Handbook. San Francisco, Calif.: American Society of Andrology.
3. Lilja H (November 2003). "Biology of prostate-specific antigen". Urology 62 (5 Suppl 1): 27–33. doi:10.1016/S0090-4295(03)00775-1.

**Plate Layout**

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