



# TF (Dog) ELISA Kit

Catalog Number KA1924

96 assays

Version: 02

Intended for research use only

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## **Introduction**

### **Intended Use**

The TF (Dog) ELISA Kit is a highly sensitive two-site enzyme linked immunoassay (ELISA) for the quantitative determination of transferrin in dog biological samples. For research use only.

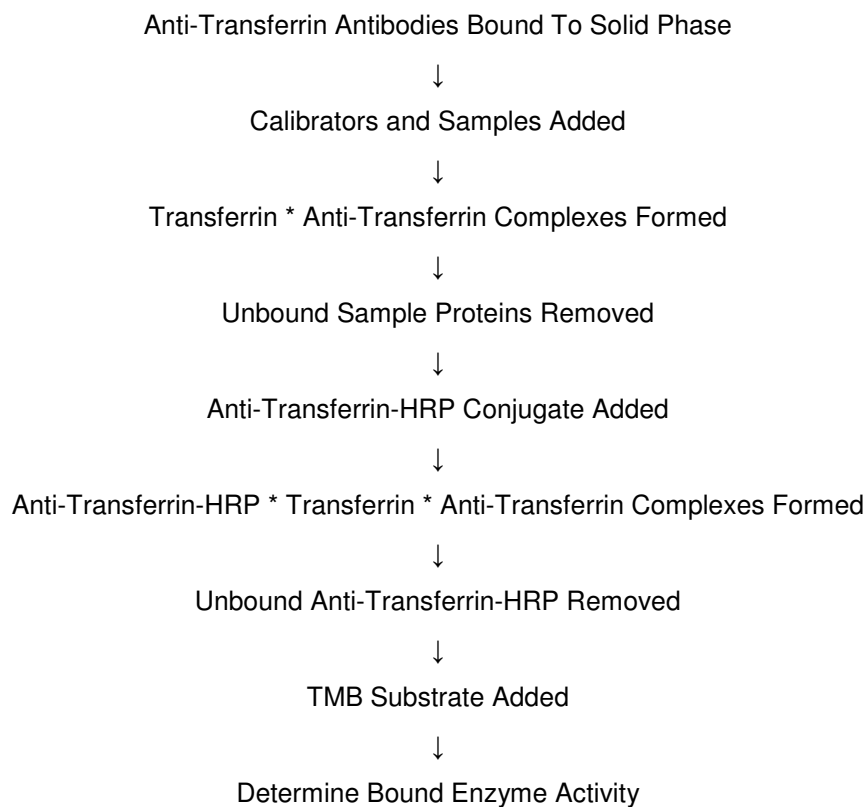
### **Background**

Transferrin (TX) is a metal-combining protein that reversibly binds to acid-soluble iron in plasma. It functions to transport iron to the bone marrow and to tissue storage organs such as the liver. Transferrin also participates in the regulation and control of iron absorption and protects against iron intoxication. Like haptoglobin, the carrier of hemoglobin, transferrin is synthesized in the liver, but unlike haptoglobin, transferrin is returned to the circulation after unloading its iron in the reticuloendothelial system. This ELISA can be used to measure transferrin in serum, tissue extracts and other biological fluids.

### **Principle of the Assay**

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the transferrin present in the sample reacts with the anti-transferrin antibodies, which have been adsorbed to the surface of polystyrene microtiter wells. After the removal of unbound proteins by washing, anti-transferrin antibodies conjugated with horseradish peroxidase (HRP), are added. This HRP-conjugated antibody forms a complex with the previously bound transferrin. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme is proportional to the concentration of transferrin in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of transferrin in the test sample. The quantity of transferrin in the test sample can be interpolated from the calibration curve constructed from the calibrators, and corrected for sample dilution.

**Figure 1.**



## General Information

### Materials Supplied

List of component

Component	Amount
Diluent Concentrate	One bottle containing 50 mL of a 5X concentrated diluent running buffer
Wash Solution Concentrate	One bottle containing 50 mL of a 20X concentrated wash solution.
Enzyme-Antibody Conjugate Concentrate	One vial containing 150 $\mu$ L of a 100X concentrated affinity-purified anti-dog transferrin antibody conjugated with HRP in a stabilizing buffer
TMB Substrate Solution	One vial containing 12 mL of TMB and hydrogen peroxide in citric acid buffer at pH 3.3
Stop Solution	One vial containing 12 mL of 0.3 M sulfuric acid. WARNING: Avoid contact with skin
Microtiter Plate	Twelve removable eight-well strips in well holder frame. Wells are coated with affinity-purified anti-dog transferrin
Dog Transferrin Calibrator	One vial containing a lyophilized Dog Transferrin Calibrator
Positive Control	One vial containing 50 $\mu$ L of serum with 0.1% sodium azide. See the Control Certificate for the concentration

### Storage Instruction

#### 1. Complete Kit

The expiration date for the kit is stated on the outer label. The recommended storage temperature is 4°C.

*Note: See long-term storage recommendations below for the Dog Transferrin Calibrator and Positive Control.*

#### 2. Diluent

The 5X Diluent Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions should be stored at 4°C.

#### 3. Wash Solution

The 20X Wash Solution Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions can be stored at room temperature (RT, 16-25°C) or at 4°C.

#### 4. Enzyme-Antibody Conjugate

Undiluted anti-transferrin-HRP conjugate should be stored at 4°C and diluted immediately prior to use. The

working conjugate solution is stable for up to 8 hours.

5. TMB Substrate Solution

The TMB Substrate Solution should be stored at 4°C and is stable until the expiration date.

6. Stop Solution

The Stop Solution should be stored at 4°C and is stable until the expiration date.

7. Microtiter Plate

Anti-dog transferrin coated wells are stable until the expiration date and should be stored at 4°C in the sealed foil pouch with a desiccant pack.

8. Dog Transferrin Calibrator

The lyophilized Dog Transferrin Calibrator should be stored at 4°C or frozen until reconstituted. The reconstituted calibrator should be aliquoted and stored frozen. Avoid multiple freeze/thaw cycles. The working calibrator solutions should be prepared immediately prior to use and are stable for up to 8 hours.

9. Positive Control

For storage longer than 7 days keep frozen until the expiration date. Storage less than 7 days can be at 4°C. Avoid multiple freeze/thaw cycles.

### **Materials Required but Not Supplied**

- ✓ Test tubes
- ✓ Precision pipette (2 µL to 200 µL) for making and dispensing dilutions
- ✓ Microplate washer/aspirator
- ✓ Distilled or de-ionized H<sub>2</sub>O
- ✓ Microplate reader
- ✓ Assorted glassware for the preparation of reagents and buffer solutions
- ✓ Timer

### **Precautions for Use**

1. Read the instructions carefully before beginning the assay.
2. This kit is for research use only.
3. Great care has been taken to ensure the quality and reliability of this product. However, it is possible that in certain cases, unusual results may be obtained due to high levels of interfering factors.
4. Preservatives: Positive Control contains 0.1% sodium azide.
5. No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.
6. Azide and thimerosal at concentrations higher than 0.1% inhibit the enzyme reaction.
7. Other precautions:

- ✓ Do not interchange kit components from different lots.
- ✓ Do not use kit components beyond the expiration date.
- ✓ Protect reagents from direct sunlight.
- ✓ Do not pipette by mouth.
- ✓ Do not eat, drink, smoke or apply cosmetics where reagents are used.
- ✓ Avoid all contact with the reagents by using gloves.
- ✓ Stop solution contains diluted sulfuric acid. Irritation to eyes and skin is possible. Flush with water after contact.

- **Limitation of the procedure**

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the instructions and with adherence to good laboratory practice.
2. Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or de-ionized water, and accuracy of reagent and sample pipettings, washing technique, incubation time or temperature.

## Assay Protocol

### Reagent Preparation

1. Diluent Concentrate

The Diluent solution supplied is a 5X concentrate and must be diluted 1:5 with distilled or de-ionized water.

2. Wash Solution Concentrate

The Wash Solution supplied is a 20X concentrate and must be diluted 1:20 with distilled or de-ionized water. Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35 °C before dilution can dissolve crystals.

3. Enzyme-Antibody Conjugate Concentrate

Calculate the required amount of working conjugate solution for each microtiter plate test strip by adding 10 µL Enzyme-Antibody Conjugate to 990 µL of 1X Diluent for each test strip to be used for testing. Mix uniformly, but gently. Avoid foaming.

4. TMB Substrate Solution

Ready to use as supplied.

5. Stop Solution

Ready to use as supplied.

6. Microtiter Plate

Ready to use as supplied. Unseal Microtiter Pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch with desiccant and re-seal.

7. Dog Transferrin Calibrator

Add 1.0 mL of distilled or de-ionized water to the lyophilized Dog Transferrin Calibrator and mix gently until dissolved. The calibrator is now at a concentration of 17.9 µg/mL (the reconstituted calibrator should be aliquoted and frozen if future use is intended). Prepare the Dog Transferrin Calibrators immediately prior to use according to the table below. Mix well between each step. Avoid foaming.

Calibrator	Concentration (ng/mL)	Calibrator Volume added to 1X Diluent	Volume of 1X Diluent
7	400	16 µL Dog Transferrin Calibrator	700 µL
6	200	250 µL Calibrator 7	250 µL
5	100	250 µL Calibrator 6	250 µL
4	50	250 µL Calibrator 5	250 µL
3	25	250 µL Calibrator 4	250 µL
2	12.5	250 µL Calibrator 3	250 µL
1	6.25	250 µL Calibrator 2	250 µL
0	0		500 µL



## 8. Positive Control

The concentration and recommended dilution are provided on the Control Certificate. Before use, briefly centrifuge the Positive Control to allow all of the liquid to collect in the bottom of the vial.

## **Sample Preparation**

Blood should be collected by venipuncture and the serum separated from the cells, after clot formation, by centrifugation. For plasma samples, blood should be collected into a container with an anticoagulant and then centrifuged. Care should be taken to minimize hemolysis, excessive hemolysis can impact your results. Assay immediately or aliquot and store samples at -20 °C. Avoid repeated freezing/thawing.

For any sample that might contain pathogens, care must be taken to prevent contact with open wounds. No additives or reservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

### • **Dilution of Samples**

The assay for quantification of Transferrin in samples requires that each test sample be diluted before use. For a single step determination, a dilution of 1:50,000 is appropriate for most serum/plasma samples. For absolute quantification of samples that yield results outside the range of the calibration curve, a lesser or greater dilution might be required. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

To prepare a 1:50,000 dilution of sample, transfer 5 µL of sample to 495 µL of 1X Diluent. This gives you a 1:100 dilution. Next, dilute the 1:100 sample by transferring 2 µL to 998 µL of 1X Diluent. You now have a 1:50,000 dilution of your sample. Mix thoroughly at each stage.

## **Assay Procedure**

1. Bring all reagents to RT before use.
2. Pipette 100 µL of
  - Calibrator 0 (0.0 ng/mL) in duplicate
  - Calibrator 1 (6.25 ng/mL) in duplicate
  - Calibrator 2 (12.5 ng/mL) in duplicate
  - Calibrator 3 (25 ng/mL) in duplicate
  - Calibrator 4 (50 ng/mL) in duplicate
  - Calibrator 5 (100 ng/mL) in duplicate
  - Calibrator 6 (200 ng/mL) in duplicate
  - Calibrator 7 (400 ng/mL) in duplicate
3. Pipette 100 µL of diluted Positive Control (in duplicate) into pre designated wells.
4. Pipette 100 µL of diluted sample (in duplicate) into pre designated wells.

5. Incubate the Microtiter Plate at 22 °C (RT) for thirty ( $30 \pm 2$ ) minutes. Keep plate covered and level during incubation.
6. Following incubation, aspirate the contents of the wells.
7. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with diluted Wash Solution, invert the plate and pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual Wash Solution. Repeat three times for a total of four washes.
8. Pipette 100  $\mu$ L of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at 22 °C (RT) for thirty ( $30 \pm 2$ ) minutes. Keep plate covered in the dark and level during incubation.
9. Wash and blot the wells as described in Steps 6 and 7.
10. Pipette 100  $\mu$ L of TMB Substrate Solution into each well.
11. Incubate in the dark at RT for precisely ten (10) minutes.
12. After ten minutes, add 100  $\mu$ L of Stop Solution to each well.
13. Determine the absorbance at 450 nm of the contents of each well. Zero the plate reader to air.

The absorbance of the final reaction mixture can be measured up to two hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.

## Data Analysis

### Calculation of Results

1. Subtract the average background value from the test values for each sample.
2. Using the results observed for the calibrators construct a calibration curve. The appropriate curve fit is that of a fourparameter logistics curve. A second order polynomial (quadratic) or other curve fits may also be used.
3. Interpolate test sample values from the calibration curve. Correct for sample dilution factor to arrive at transferring concentration in original sample.

- **Quality control**

In accord with good laboratory practice, the assays for specific transferrin require meticulous quality control. Each laboratory should use routine quality control procedures to establish inter- and intra-assay precision and performance characteristics.

- **Indications of instability**

If the test is performing correctly, the results observed with the calibrator solutions should be within 20% of the expected values.

**Resources**

**Plate Layout**

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