



# KLK1 (Human) ELISA Kit

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96 assays

Version: 06

Intended for research use only

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## **Introduction**

### **Intended Use**

For quantitative detection of human KLK1 in cell culture supernates, cell lysates, tissue homogenates, serum and plasma (heparin, EDTA).

### **Background**

Kallikrein-1, also known as tissue kallikrein, is a protein that in humans is encoded by the KLK1 gene. This serine protease generates Lys-bradykinin by specific proteolysis of kininogen-1. KLK1 is a member of the peptidase S1 family. Its gene is mapped to 19q13.3. In all, it has got 262-amino acids which contain a putative signal peptide, followed by a short activating peptide and the protease domain. The protein is mainly found in kidney, pancreas, and salivary gland, showing a unique pattern of tissue-specific expression relative to other members of the family. KLK1 is implicated in carcinogenesis and some have potential as novel cancer and other disease biomarkers.

### **Principle of the Assay**

KLK1 (Human) ELISA Kit was based on standard sandwich enzyme-linked immune-sorbent assay technology. A monoclonal antibody from mouse specific for KLK1 has been precoated onto 96-well plate. Standards (NSO, I25-S262) and the test samples are added to the wells, a biotinylated detection polyclonal antibody from goat specific for KLK1 is added subsequently and then followed by washing with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex was added and unbound conjugates were washed away with PBS or TBS buffer. HRP substrate TMB was used to visualize HRP enzymatic reaction. TMB was catalyzed by HRP to produce a blue color product that changed into yellow after adding acidic stop solution. The density of yellow is proportional to the human KLK1 amount of sample captured in plate.

## General Information

### Materials Supplied

List of component

Component	Amount
96-well plate precoated with anti- human KLK1 antibody	96 (8 x 12) wells
Lyophilized recombinant human KLK1 standard	10 ng/tube x 2
Biotinylated anti- human KLK1 antibody, dilution 1:100	130 $\mu$ L
Avidin-Biotin-Peroxidase Complex (ABC), dilution 1:100	130 $\mu$ L
Sample diluent buffer	30 mL
Antibody diluent buffer	12 mL
ABC diluent buffer	12 mL
TMB color developing agent	10 mL
TMB stop solution	10 mL

### Storage Instruction

Store at 4°C for 6 months, at -20°C for 12 months. Avoid multiple freeze-thaw cycles.

### Materials Required but Not Supplied

- ✓ Microplate reader in standard size.
- ✓ Automated plate washer.
- ✓ Adjustable pipettes and pipette tips. Multichannel pipettes are recommended in the condition of large amount of samples in the detection.
- ✓ Clean tubes and Eppendorf tubes.
- ✓ Washing buffer (neutral PBS or TBS).
- Preparation of 0.01 M TBS:  
Add 1.2 g Tris, 8.5 g NaCl; 450  $\mu$ L of purified acetic acid or 700  $\mu$ L of concentrated hydrochloric acid to 1000 mL H<sub>2</sub>O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1 L.
- Preparation of 0.01 M PBS:  
Add 8.5 g sodium chloride, 1.4 g Na<sub>2</sub>HPO<sub>4</sub> and 0.2 g NaH<sub>2</sub>PO<sub>4</sub> to 1000 mL distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1 L.

### **Precautions for Use**

- ✓ To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
- ✓ The TMB Color Developing agent is colorless and transparent before using, contact us freely if it is not the case.
- ✓ Before using the Kit, spin tubes and bring down all components to the bottom of tubes.
- ✓ Duplicate well assay is recommended for both standard and sample testing.
- ✓ Don't let 96-well plate dry, for dry plate will inactivate active components on plate.
- ✓ Don't reuse tips and tubes to avoid cross contamination.
- ✓ Avoid using the reagents from different batches together.
- ✓ In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution will be pre-warmed in 37°C for 30 min before using.

## Assay Protocol

### Reagent Preparation

- ✓ Reconstitution of the human KLK1 standard: KLK1 standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of KLK1 standard (10 ng per tube) are included in each kit. Use one tube for each experiment.
  - 10,000 pg/mL of human KLK1 standard solution: Add 1 mL sample diluent buffer into one tube, keep the tube at room temperature for 10 min and mix thoroughly.
  - 5000 pg/mL→156 pg/mL of human KLK1 standard solution: Label 6 Eppendorf tubes with 5000 pg/mL, 2500 pg/mL, 1250 pg/mL, 625 pg/mL, 312 pg/mL, 156 pg/mL respectively. Add 0.3 mL of the sample diluent buffer into each tube. Add 0.3 mL of the above 10,000 pg/mL KLK1 standard solution into 1st tube and mix. Transfer 0.3 mL from 1st tube to 2nd tube and mix. Transfer 0.3 mL from 2nd tube to 3rd tube and mix, and so on.

*Note: The standard solutions are best used within 2 hours. The 10 ng/mL standard solution may be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.*

- ✓ Preparation of biotinylated anti-human KLK1 antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.
  - The total volume should be: 0.1 mL/well x (the number of wells). (Allowing 0.1-0.2 mL more than total volume)
  - Biotinylated anti-human KLK1 antibody should be diluted in 1:100 with the antibody diluent buffer and mixed thoroughly. (i.e. Add 1 µL Biotinylated anti-human KLK1 antibody to 99 µL antibody diluent buffer.)
- ✓ Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.
  - The total volume should be: 0.1 mL/well x (the number of wells). (Allowing 0.1-0.2 mL more than total volume)
  - Avidin- Biotin-Peroxidase Complex (ABC) should be diluted in 1:100 with the ABC dilution buffer and mixed thoroughly. (i.e. Add 1 µL ABC to 99 µL ABC diluents buffer.)

### Sample Preparation

- ✓ Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

- Cell culture supernates or tissue lysates: Remove particulates by centrifugation, assay immediately or aliquot and store sample at -20°C.

- Tissue Homogenates: Rinse tissue with PBS to remove excess blood, chopped into 1-2 mm pieces, and homogenize with a tissue homogenizer in PBS or in lysate solution, lysate solution:tissue net weight = 10 mL : 1 g (i.e. Add 10 mL lysate solution to 1 g tissue). Centrifuge at approximately 5000 x g for 5 min. Assay immediately or aliquot and store homogenates at -20°C. Avoid repeated freeze-thaw cycles.
- Cell lysates: Collect cells and rinse cells with PBS. Homogenize and lyse cells thoroughly in lysate solution. Centrifuge cell lysates at approximately 10000 x g for 5 min to remove debris. Aliquot of the cell lysate were removed and assayed.
- Serum: Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 x g for 15 min. Analyze the serum immediately or aliquot and store frozen at -20°C.
- Plasma: Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 min at 1000 x g within 30 min of collection. Analyze immediately or aliquot and store frozen at -20°C.

✓ **Sample Dilution Guideline**

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. The sample must be well mixed with the diluents buffer.

- High target protein concentration (100-1000 ng/mL). The working dilution is 1:100. i.e. Add 1 µL sample into 99 µL sample diluent buffer.
- Medium target protein concentration (10-100 ng/mL). The working dilution is 1:10. i.e. Add 10 µL sample into 90 µL sample diluent buffer.
- Low target protein concentration (156-10,000 pg/mL). The working dilution is 1:2. i.e. Add 50 µL sample to 50 µL sample diluent buffer.
- Very Low target protein concentration ( $\leq 156$  pg/mL). No dilution necessary, or the working dilution is 1:2.

**Assay Procedure**

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 min before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard KLK1 detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of KLK1 amount in samples.

1. Aliquot 0.1 mL per well of the 10,000 pg/mL, 5000 pg/mL, 2500 pg/mL, 1250 pg/mL, 625 pg/mL, 312 pg/mL, 156 pg/mL human KLK1 standard solutions into the precoated 96-well plate. Add 0.1 mL of the sample diluent buffer into the control well (Zero well). Add 0.1 mL of each properly diluted sample of human cell culture supernates, cell lysates, tissue homogenates, serum or plasma (heparin, EDTA) to each empty well. See "Sample Dilution Guideline" above for details. It is recommended that each human KLK1 standard solution and each sample is measured in duplicate.

2. Seal the plate with the cover and incubate at 37°C for 90 min.
3. Remove the cover, discard plate content, and blot the plate onto paper towels or other absorbent material. Do NOT let the wells completely dry at any time.
4. Add 0.1 mL of biotinylated anti-human KLK1 antibody working solution into each well and incubate the plate at 37°C for 60 min.
5. Wash the plate 3 times with 0.01 M TBS or 0.01 M PBS, and each time let washing buffer stay in the wells for 1 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material. (Plate Washing Method: Discard the solution in the plate without touching the side walls. Blot the plate onto paper towels or other absorbent material. Soak each well with at least 0.3 mL PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of THREE washes. *Note: For automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the plate onto paper towels or other absorbent material.*)
6. Add 0.1 mL of prepared ABC working solution into each well and incubate the plate at 37°C for 30 min.
7. Wash plate 5 times with 0.01 M TBS or 0.01 M PBS, and each time let washing buffer stay in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material. (See Step 5 for plate washing method.)
8. Add 90 µL of prepared TMB color developing agent into each well and incubate plate at 37°C in dark for 20-25 min (*Note: For reference only, the optimal incubation time should be determined by end user. And the shades of blue can be seen in the wells with the four most concentrated human KLK1 standard solutions; the other wells show no obvious color.*).
9. Add 0.1 mL of prepared TMB stop solution into each well. The color changes into yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 min after adding the stop solution.

✓ Summary

1. Add samples and standards and incubate the plate at 37°C for 90 min. Do not wash.
2. Add biotinylated antibodies and incubate the plate at 37°C for 60 min. Wash plate 3 times with 0.01 M TBS.
3. Add ABC working solution and incubate the plate at 37°C for 30 min. Wash plate 5 times with 0.01 M TBS.
4. Add TMB color developing agent and incubate the plate at 37°C in dark for 20-25 min.
5. Add TMB stop solution and read.



## Data Analysis

### Calculation of Results

For calculation, (the relative O.D.<sub>450</sub>) = (the O.D.<sub>450</sub> of each well) – (the O.D.<sub>450</sub> of Zero well). The standard curve can be plotted as the relative O.D.<sub>450</sub> of each standard solution (Y) vs. the respective concentration of the standard solution (X). The human KLK1 concentration of the samples can be interpolated from the standard curve.

*Note: if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.*

✓ Typical result

Typical Data Obtained from Human KLK1

(TMB reaction incubate at 37°C for 20 min)

Concentration (pg/mL)	0.0	156	312	625	1250	2500	5000	10,000
O.D.	0.032	0.129	0.193	0.393	0.637	0.998	1.528	2.192

### Performance Characteristics

✓ Range

156 pg/mL-10,000 pg/mL

✓ Sensitivity

< 10 pg/mL

✓ Specificity

Natural and recombinant human KLK1

✓ Cross-reactivity

No detectable cross-reactivity with other relevant proteins

✓ Precision

- Intra-Assay Precision (Precision within an assay) Three samples of known concentration were tested on one plate to assess intra-assay precision.
- Inter-Assay Precision (Precision between assays) Three samples of known concentration were tested in separate assays to assess inter-assay precision.

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	16	16	16	24	24	24
Mean (pg/mL)	1468	3882	6352	1625	4190	6526
Standard deviation	67.53	260.1	349.4	95.9	301.7	417.7
CV(%)	4.6	6.7	5.5	5.9	7.2	6.4

## Resources

### References

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2. Lundwall A, Band V, Blaber M, Clements JA, Courty Y, Diamandis EP, Fritz H, Lilja H, Malm J, Maltais LJ, Olsson AY, Petraki C, Scorilas A, Sotiropoulou G, Stenman UH, Stephan C, Talieri M, Yousef GM (Jun 2006). "A comprehensive nomenclature for serine proteases with homology to tissue kallikreins". *Biol Chem* 387 (6): 637–41.
3. Diamandis, Eleftherios P.; Deperthes, David; Lundwall, Åke (Jun 2006). "Proceedings of the 1st International Symposium on Kallikreins, Lausanne, Switzerland, September 1-3, 2005". *Biol Chem* 387 (6): 635–824.

**Plate Layout**

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