

CSF3R (Human) ELISA Kit

Catalog Number KA2208

96 assays

Version: 01

Intended for research use only



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Introduction

Intended Use

Detection and Quantification of Human Granulocyte Colony-Stimulating Factor Receptor (hG-CSFR) Concentrations in Cell Lysates, Sera and Plasma.

Background

G-CSFR is a receptor for granulocyte colony-stimulating factor (CSF3), essential for granulocytic maturation. This receptor plays a crucial role in the proliferation, differentiation and survival of cells along the neutrophilic lineage. In addition, it may function in some adhesion or recognition events at the cell surface. One or several isoforms have been found in myelogenous leukemia cell line KG-1, leukemia U937 cell line, in bone marrow cells, placenta, and peripheral blood granulocytes. Isoform GCSFR-2 is found only in leukemia U937 cells while isoform GSCFR-3 is highly expressed in placenta. It has been reported that the WSXWS motif appears to be necessary for proper protein folding and thereby efficient intracellular transport and cell-surface receptor binding. Defects in CSF3R are the cause of hereditary neutrophilia, a form of lifelong, persistent neutrophilia characterized by an increase in the number of neutrophils in the blood.

Source: Entrez Gene; Swiss-Prot

Principle of the Assay

The CSF3R (Human) ELISA Kit contains the components necessary for quantitative determination of natural or recombinant hG-CSFR concentrations within any experimental sample including cell lysates, serum and plasma. This particular immunoassay utilizes the quantitative technique of a "Sandwich" Enzyme-Linked Immunosorbent Assay (ELISA) where the target protein (antigen) is bound in a "sandwich" format by the primary capture antibodies coated to each well-bottom and the secondary detection antibodies added subsequently by the investigator. The capture antibodies coated to the bottom of each well are monoclonal, specific for a particular epitope on the Human G-CSF Receptor while the user-added detection antibodies are polyclonal for binding to a variety of epitopes on the bound target protein. Amid each step of the procedure, a series of wash steps must be performed to ensure the elimination of non-specific binding between proteins to other proteins or to the solid phase. After incubation and "sandwiching" of the target antigen, a peroxidase enzyme is conjugated to the constant heavy chain of the secondary antibody (either covalently or via Avidin/Streptavidin-Biotin interactions), allowing for a colorimetric reaction to ensue upon substrate addition. When the substrate TMB (3, 3', 5, 5'-Tetramethylbenzidine) is added, the reaction catalyzed by peroxidase yields a blue color that is representative of the antigen concentration. Upon sufficient color development, the



reaction can be terminated through addition of Stop Solution (2 M Sulfuric Acid) where the color of the solution will turn yellow. The absorbance of each well can then be read by a spectrophotometer, allowing for generation of a standard curve and subsequent determination of protein concentration.



General Information

Materials Supplied

List of component

Reagent	Quantity Per Plate	Container	Reconstitution	Dilution	
96-Well Microplate					
or Strips Coated w/	-	-	-	-	
Capture Antibody					
Biotin-Conjugated				Dilute Using	
Detection Antibody	Lyophilized (1.2 µg)	Yellow	67 µl H₂O	Detection Antibody	
				Diluent	
Avidin-HRP	10 µl	Clear		Dilute Using HRP	
Conjugate	το μι	Glear	-	Diluent	
Cytokine Protein	Lyophilized (10 ng)	Red	100	Dilute Using	
Standard	Lyophilized (10 fig)	neu	100 μl H ₂ O	Standard Diluent	
Ready-to-Use	11 ml	Brown			
Substrate	111111	BIOWII	-	-	
Stop Solution	11 ml	Clear	-	-	
Adhesive Plate	4 Sheets				
Sealers	4 Sheets	•	-	-	
Wash Buffer (10X)	50 ml	Clear		Dilute to 1X Using	
	50 IIII	Glear	-	Pure H ₂ O	
Protein Standard	15 ml	Clear			
Diluent	13 1111	Gleai	-	-	
Detection Antibody	15 ml	Clear			
Diluent	13 1111	Gleai	-	-	
HRP Diluent	15 ml	Clear	-	-	

Storage Instruction

✓ Note: If used frequently, reagents may be stored at 2-8 ℃. If used infrequently, reagents should be stored at -20 ℃.

Condition	Component	Storage Information	Storage Time
Sealed, Unopened		2-8℃	1 month
Assay Kit	-	2-0 C	i inontri



Reconstituted, Opened	96-Well Microplate		
Assay Kit	(Capture Antibody Coated)		
	Detection Antibody		
	Avidin-HRP Conjugate		
	Cytokine Protein Standard		
	Ready-to-Use Substrate 2-8°C		1 month
	Stop Solution		
	Wash Buffer (10X)		
	Protein Standard Diluent		
	Detection Antibody Diluent		
	HRP Diluent		
	Plate Sealers	-	-

Materials Required but Not Supplied

- ✓ Microplate reader able to measure absorbance at 450 nm (with correction wavelength set to 540 nm or 570 nm)
- ✓ Micropipettes with capability of measuring volumes ranging from 1 µl to 1 ml
- ✓ Deionized or sterile water
- ✓ Squirt bottle, manifold dispenser, multichannel pipette reservoir or automated microplate washer
- ✓ Graph paper or computer software capable of generating or displaying logarithmic functions.
- ✓ Absorbent paper or vacuum aspirator
- ✓ Test tubes or microfuge tubes capable of storing ≥1 ml
- ✓ Bench-top centrifuge (optional)
- ✓ Bench-top vortex (optional)
- ✓ Orbital shaker (optional)

Precautions for Use

- Assay Restrictions
- ✓ This ELISA kit is intended for research purposes only, NOT diagnostic or clinical procedures of any kind.
- ✓ Materials included in this kit should NOT be used past the expiration date on the kit label.
- ✓ Reagents or substrates included in this kit should NOT be mixed or substituted with reagents or substrates from any other kits.
- ✓ Variations in pipetting technique, washing technique, operator laboratory technique, kit age, incubation time or temperature may cause differences in binding affinity of the materials provided.
- \checkmark The assay is designed to eliminate interference and background by other cellular macromolecules or



factors present within any biological samples. However, the possibility of background noise cannot be fully excluded until all factors have been tested using the assay kit.

- Health and safety Precautions
- ✓ Reagents provided in this kit may be harmful if ingested, inhaled or absorbed through the skin. Please carefully review the MSDS for each reagent before conducting the experiment.
- ✓ Stop Solution contains 2 M Sulfuric Acid (H₂SO₄) and is an extremely corrosive agent. Please wear proper eye, hand and face protection when handling this material. When the experiment is finished, be sure to rinse the plate with copious amounts of running water to dilute the Stop Solution prior to disposing the plate.



Assay Protocol

Reagent Preparation

All reagents should be diluted immediately prior to use.

Assay Procedure

- ✓ Note: If possible, all incubation steps should be performed on an orbital shaker to equilibrate solutions when added to the microplate wells. Also, all provided solutions should be at ambient temperature prior to use.
- ✓ Note: Avoid adding solutions into wells at an angle, always keep pipette tip perpendicular to plate bottom.
- Reconstitution of Provided Materials
 Please see tables above regarding reagent reconstitution and storage information.
- Addition of Known Standard and Unknown Sample to Immunoassay

Prior to applying an unknown sample to the Sandwich ELISA, the immunoassay must be performed using a serial dilution of a known standard sample in order to determine the standard curve. This is necessary to allow for the interpretation of results generated by the unknown samples.

- 1. Dilute the known standard sample from 2 ng/ml to 0 ng/ml in a series of microfuge tubes. Mix each tube thoroughly by inverting several times or by vortexing lightly to ensure proper equilibration. Add 100 µl of each serial dilution step into the wells of a specified row or column of the 96-well microtiter plate in duplicate or triplicate and incubate at room temperature for 2 hours. Seal the microplate air-tight using one of the microplate adhesive seals provided in this kit or Parafilm if readily available. *Note: If a standard curve has already been generated, substitute the standard with the unknown sample of interest.*
- Application of Detection Antibody to Capture Antibody-Bound Samples
- 1. Aspirate the protein standard solution out of the microplate wells. If your lab does not have a vacuum-based aspirator, you may dump the solutions from the microplate into a waste container and blot 3-4 times on a stack of paper towels until most or all of the liquid is removed from the wells. Dilute the 10X wash buffer to 1X using pure H₂O. Add 300-400 µl of Wash Buffer to each well being used and gently shake for 5-7 minutes on an orbital shaker. Perform this wash step 4 times consecutively.
- 2. After the 4th wash step, dilute the detection antibody solution 1:180 in detection antibody diluent to a concentration of 100 ng/ml. Mix the test tube either by inverting several times or vortexing to ensure proper equilibration. Ensure that there is enough detection antibody solution for all wells being used. Add 100 µl of the diluted detection antibody solution into each well, seal the plate and incubate at room



temperature for 2 hours.

- Conjugation of Avidin-Horseradish Peroxidase Enzyme with Detection Antibody
- 1. Remove the detection antibody solution out of the microplate wells by either vacuum-based aspirator or paper towel blotting. Perform 4 consecutive wash steps with gentle shaking between each wash.
- 2. After the 4th wash step, dilute 5.5 μl of the Avidin-HRP conjugate 1:2000 in diluent for a total volume of 11 ml. Mix the test tube either by inverting several times or vortexing to ensure proper equilibration. Add 100 μl of diluted Avidin-HRP conjugate solution into each well and incubate at room temperature for 30 minutes.
- Application of Liquid Substrate for Colorimetric Reaction
- Remove the Avidin-HRP conjugate solution out of the microplate wells by either vacuum-based aspirator
 or paper towel blotting. Prepare the TMB substrate solution by bringing it to room temperature without
 exposure to fluorescent or UV light as these may degrade the TMB. Perform 4 consecutive wash steps
 with gentle shaking between each wash.
- 2. After the 4th wash step, add 100 µl of TMB substrate solution into each well and incubate at room temperature for color development. The microplate should be kept out of direct light by either covering with an opaque object or putting it into a dark room. Closely monitor the color development as some wells may turn blue very quickly depending on analyte and/or detection antibody-HRP concentrations. Once the blue color has ceased to develop further, immediately add 100 µl of Stop Solution to each well being used. The color in the wells should immediately change from blue to yellow.
- 3. The microplate is now ready to be read by a microplate reader. Within 30 minutes of adding the Stop Solution, determine the optical density (absorbance) of each well by reading the plate with the microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. *Caution:*Readings made directly at 450 nm without correction may be higher and less accurate.



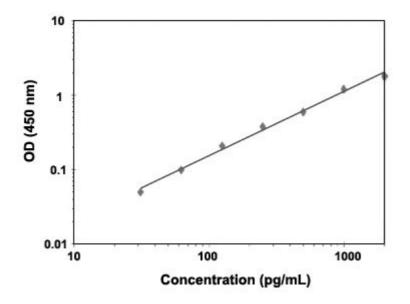
Data Analysis

Calculation of Results

- Generation of Standard Curve and Interpretation of Data
- 1. Average the duplicate or triplicate readings for each standard, control and sample and subtract the average zero standard optical density.
- 2. Generate a standard curve by using Microsoft Excel or other computer software capable of establishing a 4-Parameter Logistic (4-PL) curve fit. If using Excel or an alternative graphing tool, plot the average optical density values in absorbance units (y-axis) against the known standard concentrations in pg/ml (x-axis). Note: Only use the values in which a noticeable gradient can be established. Afterwards, generate a best fit curve or "trend-line" through the plotted points via regression analysis. Note: Shown on the next page is an example of typical data produced by analysis of the standard sample.

The data and subsequent graph was obtained after performing a cytokine ELISA for Human G-CSFR. Each known sample concentration was assayed in triplicate.

Human G-CSFR Standard Curve				
Concentration (pg/ml)	Average OD 450nm			
2000	1.8			
1000	1.2			
500	0.6			
250	0.38			
125	0.21			
62.5	0.1			
31.25	0.05			





Performance Characteristics

• Reactivity and Specificity

The CSF3R (Human) ELISA Kit is capable of recognizing both recombinant and naturally produced Human G-CSFR proteins. The antigens listed below were tested at 50 ng/ml and did not exhibit significant cross reactivity or interference.

Human: G-CSF, GM-CSF Rβ, M-CSF R/Fc Chimera

✓ Murine: G-CSF



Resources

Plate Layout

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