



IgM (Turkey) ELISA Kit

Catalog Number KA2516

96 assays

Version: 03

Intended for research use only

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Introduction

Intended Use

The IgM (Turkey) ELISA Kit is intended for measurement of IgM in serum, plasma and egg-yolk extracts.

Background

Goat anti-turkey IgM (mu chain specific) antibodies are used for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated goat anti-turkey IgM (mu chain specific) antibodies are used for detection. Normal turkey IgM levels are in the range of 2-5 mg/mL in serum or plasma (ref 1).

Principle of the Assay

Test samples are diluted and incubated in the microtiter wells for 45 minutes alongside prepared turkey IgM standards. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. IgM molecules are thus sandwiched between the immobilization and detection antibodies. The wells are then washed to remove unbound HRP-labeled antibodies and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of IgM is proportional to the optical density of the test sample and is derived from a standard curve.

General Information

Materials Supplied

List of component

Component	Amount
Anti turkey IgM coated plate	96 wells (12 strips of 8 wells)
HRP Conjugate Reagent,	11 mL
Reference Standard (lyophilized) ¹	1 vial
20x Wash Solution	50 mL
10x Immunoglobulin Diluent,	25 mL
TMB Reagent (One-step)	11 mL
Stop Solution (1 N HCl)	11 mL

¹The reference standard was calibrated with purified turkey IgM.

Storage Instruction

The test kit will remain stable for six months from the date of purchase provided that the components are stored as 2-8°C. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air.

Materials Required but Not Supplied

- ✓ Precision pipettes and tips
- ✓ Distilled or deionized water
- ✓ Polypropylene or glass tubes
- ✓ Vortex mixer
- ✓ Absorbent paper or paper towels
- ✓ Micro-Plate incubator/shaker mixing speed of ~150 rpm
- ✓ Plate washer
- ✓ Plate reader with an optical density range of 0-4 at 450nm
- ✓ Graph paper (PC graphing software is optional)

Precautions for Use

- General Instructions
- ✓ Please read and understand the instructions thoroughly before using the kit.
- ✓ All reagents should be allowed to reach room temperature (18- 25°C) before use.

- ✓ Optimum results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.
- Limitations of the procedure
- ✓ Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
- ✓ The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings. .

Assay Protocol

Reagent Preparation

- Diluent Preparation
The diluent is provided as a 10x stock. Prior to use, estimate the final volume of diluent required for your assay and dilute one (1) volume of the 10x stock with nine (9) volumes of distilled or deionized water.

- Wash Solution Preparation
The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 mL) with 950 mL of distilled or deionized water.

- Standard Preparation
 1. The IgM standard is provided as a lyophilized stock. Reconstitute with 1.0 mL of distilled or deionized water (the reconstituted standard is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended).
 2. Label 7 polypropylene or glass tubes as 200, 100, 50, 25, 12.5, 6.25, and 3.13 ng/mL.
 3. Into the tube labeled 200 ng/mL, pipette the volume of diluent detailed on the IgM standard vial label. Then add the indicated volume of IgM standard and mix gently. This provides the 200 ng/mL standard.
 4. Dispense 250 µL of diluent into the tubes labeled 100, 50, 25, 12.5, 6.25, and 3.13 ng/mL.
 5. Prepare a 100 ng/mL standard by diluting and mixing 250 µL of the 200 ng/mL standard with 250 µL of diluent in the tube labeled 100 ng/mL.
 6. Similarly prepare the 50, 25, 12.5, 6.25, 3.13 ng/mL standards by serial dilution.

Sample Preparation

General Note: In order to obtain values within range of the standard curve, we suggest that samples initially be diluted 40,000 fold using the following procedure for each sample to be tested:

- ✓ Dispense 497.5 µL of 1x diluent into two tubes.
- ✓ Pipette and mix 2.5 µL of the serum/plasma sample into the first tube and mix. This provides a 200 fold diluted sample.
- ✓ Mix 2.5 µL of the 200 fold diluted sample with the 497.5 µL of diluent in the second tube. This provides a 40,000 fold dilution of the sample.
- ✓ Repeat this procedure for each sample to be tested.

Assay Procedure

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μL of standards and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μL /well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μL of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 μL of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 20 minutes.
11. Stop the reaction by adding 100 μL of Stop Solution to each well.
12. Gently mix. It is important to make sure that all the blue color changes to yellow.
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes..

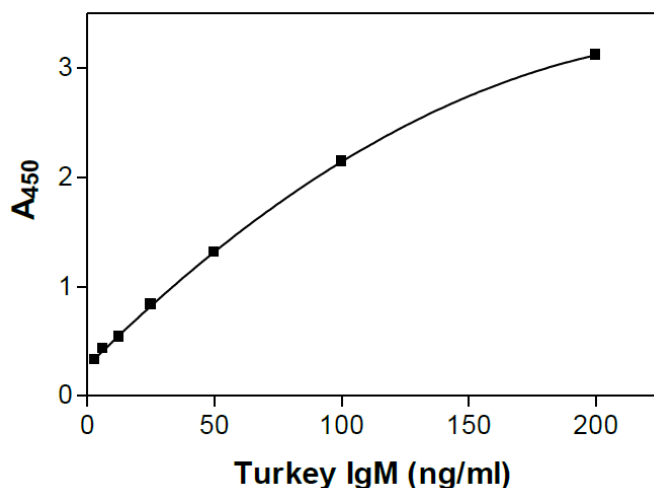
Data Analysis

Calculation of Results

1. Calculate the average absorbance values (A_{450}) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in ng/mL on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of IgM in ng/mL from the standard curve.
4. Multiply the derived concentrations by the dilution factor to determine the actual concentration of IgM in the sample.
5. PC graphing software may be used for the above steps.
6. If the OD_{450} values of samples fall outside the standard curve, samples should be diluted appropriately and re-tested.

A typical standard curve with optical density readings at 450 nm on the Y axis against IgM concentrations on the X axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

IgM (ng/mL)	Absorbance (450 nm)
200	3.120
100	2.146
50	1.311
25	0.833
12.5	0.540
6.25	0.432
3.13	0.330



Resources

Reference

1. Li Z, Nestor KE, Saif YM, Anderson JW and Patterson RA. Serum immunoglobulin G and M concentrations did not appear to be associated with resistance to *Pasteurella multocida* in a large-bodied turkey line and a random population control. *Poultry Science* 79: 163-166 (2000)

Plate Layout

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