



PRODUCT INFORMATION & MANUAL

DNA Damage Quantification Assay Kit (Colorimetric)

NBP2-54878

For research use only.

Not for diagnostic or therapeutic procedures.

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Novus kits are guaranteed for 6 months from date of receipt

DNA Damage Quantification Colorimetric Kit

(Catalog #NBP2-54878; Store kit at 4°C)

I. Introduction:

Apurinic/aprimidinic (AP) sites are one of the major types of DNA lesions formed during the course of base excision and repair of oxidized, deaminated or alkylated bases. It has been estimated that about 2×10^5 base lesions are generated per cell per day. The level of AP sites in cells can be a good indicator of DNA lesion and repair against chemical damage and cell aging. The *DNA Damage Quantification Kit* utilizes the ARP (Aldehyde Reactive Probe) reagent that reacts specifically with an aldehyde group which is the open ring form of the AP sites. After treating DNA containing AP sites with ARP reagents, AP sites are tagged with biotin residues, which can be quantified using avidin-biotin assay followed by a colorimetric detection. The kit provides the necessary reagents for convenient determination of abasic sites in purified DNA sample in 96-well plate format.

II. Kit Contents:

Component	NBP2-54878	Color Code
	25 assays	Cap Color
ARP Solution (10 mM, in DMSO)	0.125 ml	Red
TE Buffer	30 ml	NM
Glycogen Solution (10 µg/µl)	0.1 ml	Blue
0 ARP-DNA Standard (0.5 µg/ml)	0.6 ml	Clear
40 ARP-DNA Standard (0.5 µg/ml)	0.6 ml	Yellow
DNA Binding Solution	10 ml	WM
HRP-Streptavidin	0.1 ml	Green
10X Wash Buffer	30 ml	WM
HRP Developer	10 ml	Brown/NM
96-well Microplate (8 x 12 strips)	1 plate	Clear

III. DNA Damage Quantification Protocol:

A. Purification of Genomic DNA:

Several different methods and products are available for isolating genomic DNA. Among all the methods, the guanidine/detergent lysis method is simple, and it gives highly purified genomic DNA for the ARP-based abasic sites detection. During the purification process, avoid heating of the DNA solution. Determine the concentration and purity of the purified genomic DNA using the spectrophotometer*. Dissolve the genomic DNA in TE at concentration of 0.1 µg/µl. It is important for an accurate assay that the DNA concentration is adjusted exactly to 0.1 µg/µl.

$1.0 \text{ OD}_{260 \text{ nm}} = 50 \text{ µg/ml}$ for genomic DNA. The ratio of $\text{OD}_{260 \text{ nm}}/\text{OD}_{280 \text{ nm}}$ of highly purified DNA solution is 1.7 or higher. Protein contamination in the sample solution may cause a positive error.

B. ARP Labeling:

- 1) Mix 5 µl of 0.1 µg/µl purified sample DNA solution with 5 µl ARP Solution at the bottom of a microcentrifuge tube and incubate at 37°C for 1 hour to tag the DNA AP site.
- 2) Add 88 µl TE and 2 µl Glycogen to the reaction solution, mix well.
- 3) Add 0.3 ml of pure ethanol (not provided) mix well and keep at -20°C for 10 min. Centrifuge with microcentrifuge at the top speed for 10 min to precipitate the AP-site tagged DNA.
- 4) Wash the pellet three times with 0.5 ml 70 % ethanol. Quick spin to remove the trace amount of ethanol. Air dry the pellet for 5 min. The Biotin-tagged genomic DNA pellet can be used immediately or store at -20°C. The tagged DNA sample is stable for at least one year.

C. Determination of the number of abasic sites in DNA

- 1) Dilute the 40 ARP-DNA Standard (40 ARP sites per 10^5 bp) with 0 APR-DNA Standard to generate 200 µl each of the 0, 8, 16, 24, 32, 40 ARP-DNA solutions in microcentrifuge tubes (see below table).

ARP Number	0	8	16	24	32	40
40 ARP-DNA Standard (µl)	0	40	80	120	160	200
0 ARP-DNA Standard (µl)	200	160	120	80	40	0

- 2) Dissolve the Biotin-tagged DNA samples prepared in B with 1 ml of TE buffer (0.5 µg/ml).
- 3) Add 60 µl each of the above ARP-DNA Standards and ARP-labeled DNA samples into each well. For more accurate measurement, use three wells per sample.
- 4) Add 100 µl of the DNA Binding Solution to the standards and samples, keep the plate at room temperature overnight to allow the tagged-DNA bind on the plate surface. Keep the wells sealed.

Prepare Solutions before use:

Washing Buffer: Dilute the 10X Wash Buffer to 1X Buffer with ddH₂O (total volume 300 ml). Store this 1X Wash Buffer at room temperature.

HRP-Streptavidin Solution: Just before use, prepare 1:100 diluted working solution by diluting the 100 µl of HRP-Streptavidin with 9.9 ml 1X Wash Buffer.

- 5) Discard the DNA Binding Solution in the wells, and wash the well with 250 µl Wash Buffer 5 times.
- 6) Add 100 µl diluted HRP-Streptavidin solution to each well, and shake the plate for 1 hr at room temperature.
- 7) Discard the solution in the wells, and wash the wells with 250 µl Wash Buffer 5 times.
- 8) Add 100 µl of HRP Developer to each well, and incubate at 37°C for 1 hour.
- 9) Measure the OD 650 nm (within 1 hour after the incubation is ended), or add 100 µl of 1 M Sulfuric acid (or 6 M HCl) to stop the reaction. Mix well and measure OD 450 nm.*

***Note:** The value from OD 450 nm will be roughly double of that from OD 650 nm.

- 10) Prepare the calibration curve using the data obtained with standard ARP-DNA solutions. Apply your sample DNA readings to the Standard Curve. The ARP numbers are the basic sites per 10^5 bp in the genomic DNA samples. Compare the numbers of AP sites in treated samples vs control samples to determine the level of DNA Damage.