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# PRODUCT INFORMATION & ELISA MANUAL

# Complement Component C2 Antibody Pair [HRP] NBP2-79453

# Sample Insert for reference use only

Matched Antibody Pair utilized in an Enzymelinked Immunosorbent Assay for quantitative detection of Human Complement Component C2.

For research use only. Not for diagnostic or therapeutic procedures.

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Novus kits are guaranteed for 6 months from date of receipt

## BACKGROUND

Complement component C2 is part of the classical complement pathway which plays a major role in innate immunity against infection. C2 is a glycoprotein synthesized in liver hepatocytes and several other cell types in extrahepatic tissues. This pathway is triggered by a multimolecular complex C1, and subsequently the single-chain form of C2 is cleaved into two chains referred to C2a and C2b by activated C1. The second component of complement (C2) is a multi-domain serine protease that provides catalytic activity for the C3 and C5 convertases of the classical and lectin pathways of human complement. C4b and C2 was investigated by surface plasmon resonance. C2a containing a serine protease domain combines with complement component C4b to form the C3 convertase C4b2a which is responsible for C3 activation, and leads to the stimulation of adaptive immune responses via Lectin pathway. C2 bound to C4b is cleaved by classical (C1s) or lectin (MASP2) proteases to produce C4bC2a. C2 has the same serine protease domain as C4bC2a but in an inactive zymogen-like conformation, requiring cofactor-induced conformational change for activity. Deficiency of C2 (C2D) is the most common genetic deficiency of the complement system, and two types of C2D have been recognized in the context of specific MHC haplotypes. C2D in human is reported to increase susceptibility to infection, and is associated with certain autoimmune diseases, such as rheumatological disorders.

## PRINCIPLE OF THE TEST

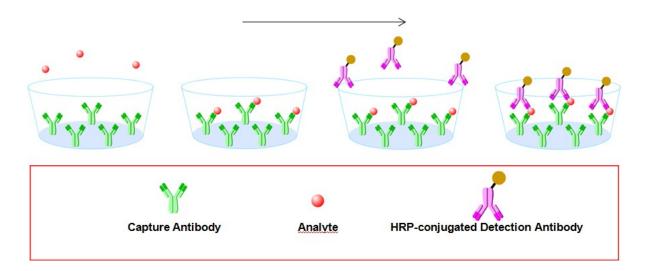
The Novus Biologicals Complement Component C2 Antibody Pair [HRP] is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for Complement Component C2 coated on a 96-well plate. Standards and samples are added to the wells, and any Complement Component C2 present binds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated mouse anti- Complement Component C2 monoclonal antibody is then added, producing an antibody-antigen-antibody "sandwich". The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of Complement Component C2 present in the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

#### INTENDED USE

◆ The Human Complement Component C2 Antibody Pair [HRP] is for the quantitative determination of Human Complement Component C2.

 This Complement Component C2 Antibody Pair [HRP] contains the basic components required for the development of sandwich ELISAs.

#### ASSAY PROCEDURE SUMMARY



This antibody pair has been configured for research use only and is not to be used in diagnostic procedures.

# MATERIALS PROVIDED

Bring all reagents to room temperature before use.

**Capture Antibody** – 0.5 mg/mL of rabbit anti-Complement Component C2 monoclonal antibody. Dilute to a working concentration of 2.0 µg/mL in CBS before coating.

**Detection Antibody** – 0.5 mg/mL mouse anti-Complement Component C2 monoclonal antibody conjugated to horseradish-peroxidase (HRP). Dilute to working concentration of 1  $\mu$ g/mL in detection antibody dilution buffer before use.

**Standard** – Each vial contains 240 ng of recombinant Complement Component C2. Reconstitute standard powder with 1 mL detection antibody dilution buffer. After reconstitution, store at  $-20^{\circ}$ C to  $-80^{\circ}$ C in a manual defrost freezer. A seven-point standard curve using 2-fold serial dilutions in sample dilution buffer, and a high standard of 5000 pg/mL is recommended.

# SOLUTIONS REQUIRED

CBS - 0.05M Na<sub>2</sub>CO<sub>3</sub>, 0.05M NaHCO<sub>3</sub>, pH 9.6, 0.2 µm filtered

**TBS** - 20 mM Tris, 150 mM NaCl, pH 7.4

Wash Buffer - 0.05% Tween20 in TBS, pH 7.2 - 7.4

Blocking Buffer - 2% BSA in Wash Buffer

Sample dilution buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

**Detection antibody dilution buffer** - 0.5% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Substrate Solution : To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10mg / ml TMB ( Tetramethylbenzidine ) in DMSO

Substrate dilution buffer -  $0.05M Na_2HPO_4$  and 0.025M citric acid; adjust pH to 5.5

Substrate working solution - For each plate dilute 250 µl substrate stock solution in 25ml

substrate dilution buffer and then add 80  $\mu$ I 0.75% H<sub>2</sub>O<sub>2</sub>, mix it well

Stop Solution - 2 N H<sub>2</sub>SO<sub>4</sub>

# PRECAUTION

The Stop Solution suggested for use with this antibody pair is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

# STORAGE

**Capture Antibody**: Aliquot and store at -20  $^{\circ}$ C to -80  $^{\circ}$ C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody**: Protect it from prolonged exposure to light. Aliquot and store at -20  $^{\circ}$ C to - 80  $^{\circ}$ C and for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Standard**: Store lyophilized standard at -20  $^{\circ}$ C to -80  $^{\circ}$ C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at -80  $^{\circ}$ C for up to 1 month. Avoid repeated freeze-thaw cycles.

## **GENERAL ELISA PROTOCOL**

#### Plate Preparation

1. Dilute the capture antibody to the working concentration in CBS. Immediately coat a 96-well microplate with 100 $\mu$ L per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.

2. Aspirate each well and wash with at least 300µl wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels. 3.Block plates by adding 300 µL of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.

4.Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

#### Assay Procedure

1.Add 100  $\mu$ L of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.

2. Repeat the aspiration/wash as in step 2 of plate preparation.

3. Add 100 µL of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.

4. Repeat the aspiration/wash as in step 2 of plate preparation.

5.Add 200  $\mu$ L of substrate solution to each well. Incubate for 20 minutes at room temperature ( **if substrate solution is not as requested, the incubation time should be optimized**). Avoid placing the plate in direct light.

6. Add 50  $\mu$ L of stop solution to each well. Gently tap the plate to ensure thorough mixing. 7. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

## CALCULATION OF RESULTS

• Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.

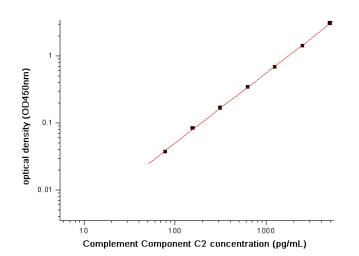
• Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.

•To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the yaxis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

•Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

#### **TYPICAL DATA**

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



Concentration (pg/ml)	Zero standard subtracted OD
0	0.000
78.125	0.037
156.25	0.084
312.5	0.168
625	0.345
1250	0.686
2500	1.412
5000	3.064

#### PERFORMANCE CHARACTERISTIC

#### SENSITIVITY

The minimum detectable dose of Human Complement Component C2 was determined to be approximately 78.125 pg/ml. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

# **TROUBLE SHOOTING**

Problems	Possible Sources	Solutions			
	Incorrect or no Detection Antibody was added	Add appropriate Detection Antibody and continue			
No signal	Substrate solution was not added	Add substrate solution and continue			
	Incorrect storage condition	Check if the kit is stored at recommended condition and used before expiration date			
	Standard was incompletely reconstituted or was inappropriately stored	Aliquot reconstituted standard and store at -80 $^\circ\!\!\!\!C$			
Poor Standard	Imprecise / inaccurate pipetting	Check / calibrate pipettes			
Curve	Incubations done at inappropriate temperature, timing or agitation	Follow the general ELISA protocol			
	Background wells were contaminated	Avoid cross contamination by using the sealer appropriately			
	The concentration of antigen in samples was too low	Enriching samples to increase the concentration of antigen			
Poor detection value	Samples were ineffective	Check if the samples are stored at cold environment. Detect samples in timely manner			
	Insufficient washes	Use multichannel pipettes without touching the reagents on the plate			
High Background	insuncient wasnes	Increase cycles of washes and soaking time between washes			
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells			
	Materials were contaminated.	Use clean plates, tubes and pipettes tips			
Non-specificity	Samples were contaminated	Avoid cross contamination of samples			
	The concentration of samples was too high	Try higher dilution rate of samples			

	ELISA Plate Template											
	1	2	3	4	5	6	7	8	9	10	11	12
Α												
В												
С												
D												
E												
F												
G												
H												

# Human Complement Component C2 Antibody Pair [HRP] Notes