

# Product Information & ELISA Manual

## LAMP-1/CD107a Antibody Pair [HRP] NBP2-79616

Matched Antibody Pair utilized in an Enzyme-linked Immunosorbent Assay for quantitative detection of Human LAMP-1/CD107a.

### Sample Insert for Reference Use Only

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Novus kits are  
guaranteed for 6 months  
from date of receipt.

**For research use only.  
Not for diagnostic or  
therapeutic procedures.**

## BACKGROUND

Lysosome-associated membrane glycoprotein 1, also known as CD107 antigen-like family member A, CD107a, and LAMP1, is a single-pass type I membrane protein that belongs to the LAMP family. CD107a is expressed largely in the endosome-lysosome membranes of cells but is also found on the plasma membrane (1-2% of total LAMP1). LAMP1 has been implicated in a variety of cellular functions, including cancer metastasis. It has been proposed LAMP1 serves as a therapeutic agent for some cancers, as well as a marker for lysosomal storage disorders and different cell types such as cytotoxic T cells. LAMP2, also known as CD107b, may also play a role in tumor cell metastasis and functions in the protection, maintenance, and adhesion of the lysosome. Cell surface LAMP1 and LAMP2 have been shown to promote adhesion of human peripheral blood mononuclear cells (PBMC) to vascular endothelium, therefore they are possibly involved in the adhesion of PBMCs to the site of inflammation. LAMP-1 is a glycoprotein highly expressed in lysosomal membranes. The present study was initiated to test LAMP-1 mRNA and protein levels in post mortem frontal cortex (area 8) of Alzheimer's disease (AD) stages I-IIA/B and stages V-VIC of Braak and Braak, compared with age-matched controls. LAMP-1 occurred in microglia and multinucleated giant cells in one AD case in which amyloid burden was cleared following beta A-peptide immunization. Also, LAMP-1 has been suggested to be a cell surface receptor for a specific amelogenin isoform, leucine-rich amelogenin peptide, or LRAP. LAMP-1 can serve as a cell surface binding site for amelogenin on dental follicle cells and cementoblasts.

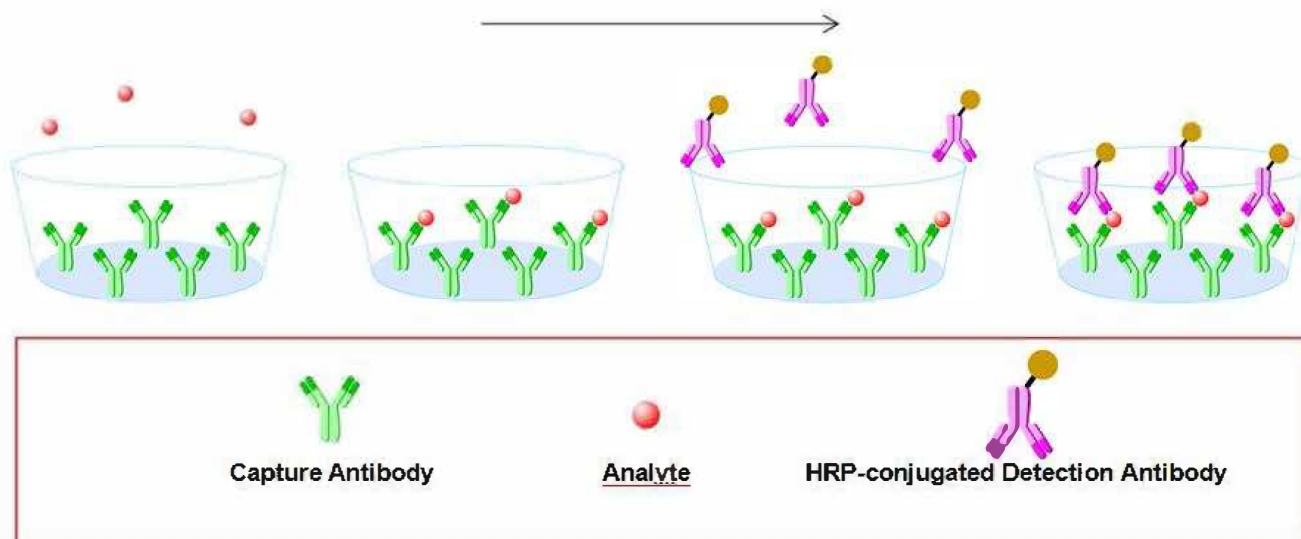
## PRINCIPLE OF THE TEST

This ELISA Pair Set contains the basic components required for the development of sandwich ELISAs. The Novus Biologicals ELISA Pair Set is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for LAMP-1/CD107a coated on a 96-well plate. Standards and samples are added to the wells, and any LAMP-1/CD107a present binds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated mouse anti-LAMP-1/CD107a monoclonal antibody is then added, producing an antibody-antigen-antibody “sandwich”. The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of LAMP-1/CD107a present in the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

## INTENDED USE

- ◆ The Human LAMP-1/CD107a ELISA Pair Set is for the quantitative determination of Human LAMP-1/CD107a.
- ◆ This ELISA Pair Set contains the basic components required for the development of sandwich ELISAs.

## ASSAY PROCEDURE SUMMARY



**This Pair Set has been configured for research use only and is not to be used in diagnostic procedures.**

## MATERIALS PROVIDED

**Bring all reagents to room temperature before use.**

**Capture Antibody** – 1 mg/mL of mouse anti-LAMP1 monoclonal antibody (in PBS, pH 7.4). Dilute to a working concentration of 2 µg/mL in PBS before coating.

**Detection Antibody** – 0.2 mg/mL mouse anti-LAMP1 monoclonal antibody conjugated to horseradish-peroxidase (HRP) (in PBS, 50 % HRP-Protector, pH 7.4, store at 4°C). Dilute to working concentration of 0.4 µg/mL in dilution buffer before use.

**Standard** – Each vial contains 42 ng of recombinant LAMP1. Reconstitute with 1 mL dilution buffer. After reconstitution, store at -20°C to -80°C in a manual defrost freezer. A seven-point standard curve using 2-fold serial dilutions in dilution buffer, and a high standard of 1000 pg/mL is recommended.

**Standard reconstitution tips:** *Add dilution buffer, gently mix it up and down 3~5 times. Avoid violent and long-time shock.*

## SOLUTIONS REQUIRED

**PBS** - 136.9 mM NaCl, 10.1 mM Na<sub>2</sub>HPO<sub>4</sub>, 2.7 mM KCl, 1.8 mM KH<sub>2</sub>PO<sub>4</sub>, pH 7.4, 0.2 µm filtered

**Wash Buffer** - 0.05% Tween20 in PBS, pH 7.2 - 7.4

**Blocking Buffer** - 2% BSA in Wash Buffer

**Dilution Buffer** - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

**Substrate Solution** : To achieve best assay results, fresh substrate solution is recommended

**Substrate stock solution** - 10mg / ml TMB ( Tetramethylbenzidine ) in DMSO

**Substrate dilution buffer** - 0.05M Na<sub>2</sub>HPO<sub>4</sub> and 0.025M citric acid ; adjust pH to 5.5

**Substrate working solution** - For each plate dilute 250 µl substrate stock solution in 25ml substrate dilution buffer and then add 80 µl 0.75% H<sub>2</sub>O<sub>2</sub> , mix it well

**Stop Solution** - 2 N H<sub>2</sub>SO<sub>4</sub>

## PRECAUTION

The Stop Solution suggested for use with this Pair Set is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

## STORAGE

**Capture Antibody:** Aliquot and store at  $-20^{\circ}\text{C}$  to  $-80^{\circ}\text{C}$  for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody:** Store at  $4^{\circ}\text{C}$  and protect it from prolonged exposure to light for up to 6 months from date of receipt. **DO NOT FREEZE!**

**Standard:** Store lyophilized standard at  $-20^{\circ}\text{C}$  to  $-80^{\circ}\text{C}$  for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at  $-80^{\circ}\text{C}$  for up to 1 month. Avoid repeated freeze-thaw cycles.

## GENERAL ELISA PROTOCOL

### Plate Preparation

1. Dilute the capture antibody to the working concentration in PBS. Immediately coat a 96-well microplate with 100µL per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
2. Aspirate each well and wash with at least 300µl wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels.
3. Block plates by adding 300 µL of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

### Assay Procedure

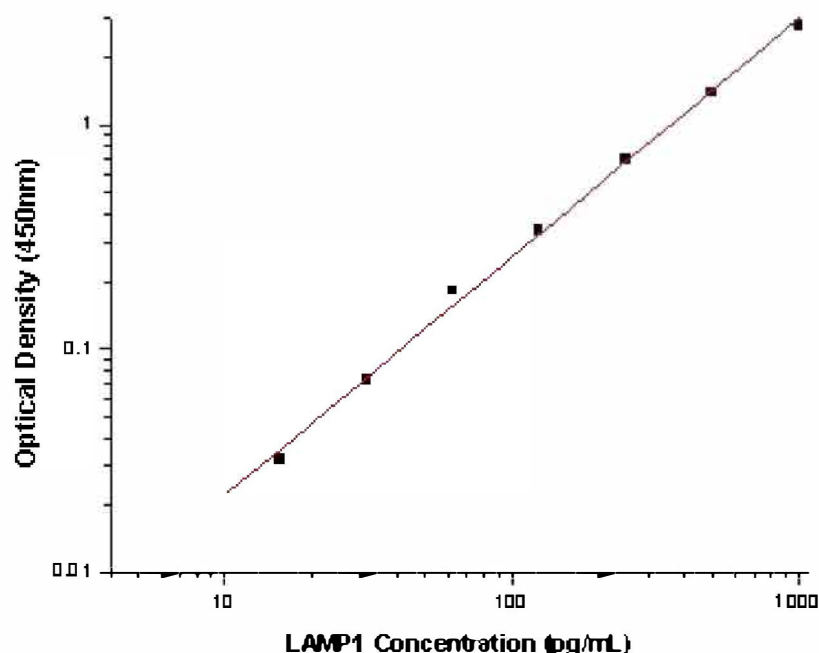
1. Add 100 µL of sample or standards in Dilution Buffer per well. Seal the plate and incubate 2 hours at room temperature.
2. Repeat the aspiration/wash as in step 2 of plate preparation.
3. Add 100 µL of the detection antibody, diluted in Dilution Buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
4. Repeat the aspiration/wash as in step 2 of plate preparation.
5. Add 200 µL of substrate solution to each well. Incubate for 20 minutes at room temperature ( **if substrate solution is not as requested, the incubation time should be optimized** ). Avoid placing the plate in direct light.
6. Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
7. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

## CALCULATION OF RESULTS

- Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

## TYPICAL DATA

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



| Concentration ( pg/mL) | Zero standard subtracted OD |
|------------------------|-----------------------------|
| 0                      | 0                           |
| 15.63                  | 0.032                       |
| 31.25                  | 0.072                       |
| 62.5                   | 0.181                       |
| 125                    | 0.339                       |
| 250                    | 0.699                       |
| 500                    | 1.387                       |
| 1000                   | 2.773                       |

## PERFORMANCE CHARACTERISTIC

### SENSITIVITY

The minimum detectable dose of Human LAMP-1/CD107a was determined to be approximately 15.63 pg/mL. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

## TROUBLE SHOOTING

| Problems                    | Possible Sources  | Solutions  |
|-----------------------------|---|--|
| <b>No signal</b>            | Incorrect or no Detection Antibody was added                          | Add appropriate Detection Antibody and continue                                      |
|                             | Substrate solution was not added                                      | Add substrate solution and continue  |
|                             | Incorrect storage condition   | Check if the kit is stored at recommended condition and used before expiration date  |
| <b>Poor Standard Curve</b>  | Standard was incompletely reconstituted or was inappropriately stored | Aliquot reconstituted standard and store at -80 °C                                   |
|                             | Imprecise / inaccurate pipetting                                      | Check / calibrate pipettes   |
|                             | Incubations done at inappropriate temperature, timing or agitation    | Follow the general ELISA protocol  |
|                             | Background wells were contaminated                                    | Avoid cross contamination by using the sealer appropriately                          |
| <b>Poor detection value</b> | The concentration of antigen in samples was too low                   | Enriching samples to increase the concentration of antigen                           |
|                             | Samples were ineffective  | Check if the samples are stored at cold environment. Detect samples in timely manner |
| <b>High Background</b>      | Insufficient washes   | Use multichannel pipettes without touching the reagents on the plate                 |
|                             |   | Increase cycles of washes and soaking time between washes                            |
|                             | TMB Substrate Solution was contaminated                               | TMB Substrate Solution should be clear and colorless prior to addition to wells      |
|                             | Materials were contaminated.  | Use clean plates, tubes and pipettes tips  |
| <b>Non-specificity</b>      | Samples were contaminated   | Avoid cross contamination of samples   |
|                             | The concentration of samples was too high                             | Try higher dilution rate of samples  |

# ELISA Plate Template

[illegible]

# **Human LAMP-1/CD107a ELISA Pair Set**

## **Notes**