



PRODUCT INFORMATION & MANUAL

Hydrogen Peroxide Assay Kit (Colorimetric) *NBP3-24506*

For research use only.
Not for diagnostic or therapeutic
procedures.

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Hydrogen Peroxide Assay Kit (Colorimetric)

Catalog No: NBP3-24506

Method: Colorimetric method

Specification: 100 Assays (Can detect 96 samples without duplication)

Instrument: Spectrophotometer

Sensitivity: 1.5 mmol/L

Detection range: 1.5-150 mmol/L

Average intra-assay CV (%): 1.3

Average inter-assay CV (%): 2.7

Average recovery rate (%): 98

- ▲ This kit is for research use only.
- ▲ Instructions should be followed strictly, changes of operation may result in unreliable results.
- ▲ Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

General information

▲ Intended use

This kit can be used to measure the H_2O_2 content in serum, plasma, tissue, cells and culture supernatant samples.

▲ Background

Hydrogen peroxide (H_2O_2) is a metabolic by-product of reactive oxygen species, which is not only a signal molecule in cells, but also a source of oxidative stress. H_2O_2 is an important regulatory factor of eukaryotic signal transduction involved in cell proliferation, differentiation and migration. However, abnormal H_2O_2 can lead to oxidative cell damage and disease, such as cancer, atherosclerosis, osteoporosis and neurodegenerative diseases.

▲ Detection principle

Hydrogen peroxide can react with ammonium molybdate to form a yellow complex which has a maximum absorption peak at 405 nm. H_2O_2 content can be calculated by measuring the absorbance value at 405 nm.

▲ **Kit components & storage**

Item	Component	Specification	Storage
Reagent 1	Buffer Solution	60 mL × 2 vials	2-8℃ , 12 months
Reagent 2	Ammonium Molybdate Reagent	60 mL × 2 vials	2-8℃ , 12 months
Reagent 3	1 mol/L H ₂ O ₂ Standard	12 mL × 1 vial	2-8℃ , 12 months
Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.			

▲ **Materials prepared by users**

 Instruments

Spectrophotometer (405 nm), Micropipettor, Incubator, Vortex mixer, Centrifuge

 Reagents

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

▲ Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes.

▲ The key points of the assay

1. Preheat the reagent 1 at 37 °C for 10 minutes. Dissolve the reagent 2 in 60 °C water bath if crystallized.
2. If the concentration of H_2O_2 in the sample is too high, please dilute the samples appropriately. If the concentration is too low, the sampling volume of the sample should be increased, and the sampling volume of standard and double distilled water should be increased equally at the same time.

Pre-assay preparation

▲ Reagent preparation

1. Preheat reagent 1 in 37°C for 10 min before use.
2. Reagent 2 is supersaturated solution. Incubate it in 60°C water bath if crystallized.
3. Preparation of 60 mmol/L H_2O_2 standard solution
Prepare fresh solution by diluting the 1 mol/L H_2O_2 standard stock solution with double distilled water at a ratio of 3:47 before use.

▲ **Sample preparation**

The samples should be prepared as conventional methods. Also please refer to appendix II.

▲ **Dilution of sample**

It is recommended to take 2~3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (1.5-150 mmol/L).

The recommended dilution factor for different samples is as follows (for reference only)

Sample type	Dilution factor
10% Mouse liver tissue homogenization	5-10
Human serum	1
10% Green pepper tissue homogenization	1
Rat serum	1
Mouse serum	1

Note: The diluent is normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

Assay protocol

▲ Detailed operation steps

1. Add 1 mL of reagent 1 to 5 mL EP tubes and incubate the tubes in 37°C for 10 min.
2. **Blank tube:** Add 0.1 mL of double distilled water to the tube.
Standard tube: Add 0.1 mL of 60 mmol/L H₂O₂ standard solution to the tube.
Sample tube: Add 0.1 mL of sample to the tube.
3. Add 1 mL of reagent 2 to each tube of Step 2 and mix fully.
4. Set the spectrophotometer to zero with double distilled water, then measure the OD value of each tube at 405 nm with 1 cm optical path quartz cuvette.

▲ Summary operation table

	Blank tube	Standard tube	Sample tube
Reagent 1 (mL)	1	1	1
Preheat at 37°C for 10 min			
Double distilled water (mL)	0.1		
60 mmol/L H ₂ O ₂ standard solution (mL)		0.1	
Sample (mL)			0.1
Reagent 2 (mL)	1	1	1
Mix fully, set the spectrophotometer to zero, then measure the OD value of each tube.			

▲ Calculation

1. Serum (plasma) and other liquid sample:

$$\text{H}_2\text{O}_2 \text{ content (mmol/L)} = \frac{\Delta A_1}{\Delta A_2} \times c \times f$$

2. Tissue and cells sample:

$$\text{H}_2\text{O}_2 \text{ content (mmol/gprot)} = \frac{\Delta A_1}{\Delta A_2} \times c \times f \div C_{\text{pr}}$$

Note:

ΔA_1 : $\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}$

ΔA_2 : $\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}$

c: The concentration of H_2O_2 standard, 60 mmol/L.

f: The dilution factor of sample before test.

C_{pr} : The concentration of protein in sample, gprot/L

Appendix I Data

▲ Example analysis

Take 0.1 mL of human serum, carry the assay according to the operation table.

The results are as follows:

The average OD value of the sample is 0.445, the average OD value of the blank is 0.051, the average OD value of the standard is 0.422, and the calculation result is:

$$\text{H}_2\text{O}_2 \text{ content (mmol/L)} = \frac{0.445-0.051}{0.422-0.051} \times 60 \times 1 = 63.72 \text{ mmol/L}$$

Appendix II Sample preparation

The following sample pretreatment methods are for reference only.

▲ Serum

Collect fresh blood and stand at 25°C for 30 min to clot the blood. Then centrifuge at 2000 g for 15 min at 4°C . Take the serum (which is the upper light yellow clarified liquid layer) to preserve it on ice for detection. If not detected on the same day, the serum can be stored at -80°C for a month.

▲ Plasma

Take fresh blood into the tube which has anticoagulant, centrifuge at 700-1000 g for 10 min at 4°C . Take the plasma (which is the upper light yellow clarified liquid layer, don't take white blood cells and platelets in the middle layer) to preserve it on ice for detection. If not detected on the same day, the plasma can be stored at -80°C for a month.

▲ Cell culture supernatant

Detect directly. If there is turbidity, centrifuge at 3100 g for 10 min. Take the supernatant to preserve it on ice for detection. If not detected on the same day, it can be stored at -80°C for a month.

▲ Tissue sample

Take 0.02-1g fresh tissue to wash with PBS (0.01 M, pH 7.4) at 2-8°C . Absorb the water with filter paper and weigh. Homogenize at the ratio of the volume of homogenized medium (2-8°C) (mL): the weight of the tissue (g) =9:1, then centrifuge the tissue homogenate for 10 min at 10000 g at 4°C . Take the supernatant to preserve it on ice for detection. Meanwhile, determine the protein concentration of supernatant. If not detected on the same day, the tissue sample (without homogenization) can be stored at -80°C for a month.

▲ Cells

Collect the cells and wash the cells with PBS (0.01 M, pH 7.4) for 1~2 times. Centrifuge at 1000 g for 10 min and then discard the supernatant and keep the cell sediment. Add homogenization medium at a ratio of cell number (10^6): homogenization medium (μL) = 1: 300-500. Sonicate or grind with hand-operated in ice water bath. Centrifuge at 10000 g for 10 min, then take the supernatant and preserve it on ice for detection. Meanwhile, determine the protein concentration of supernatant. If not detected on the same day, the cells sample (without homogenization) can be stored at -80°C for a month.

Note:

1. Homogenized medium: PBS (0.01 M, pH 7.4) including 0.1 mM EDTA.
2. Homogenized method:
 - (1) Hand-operated: Weigh the tissue and mince to small pieces (1 mm^3), then put the tissues pieces to glass homogenized tube. Add homogenized medium into homogenized tube, place the tube into the ice bath with left hand, and insert the glass tamping rod vertically into the homogenized tube with the right hand to grind up and down for 6-8 min.

Or put the tissue into the mortar, and add liquid nitrogen to grind fully. Then add the homogenized medium to homogenize.
 - (2) Mechanical homogenate: Weigh the tissue to EP tube, add the homogenized medium to homogenize the tissue with homogenizer instrument (60 Hz, 90s) in the ice bath. (For samples of skin, muscle and plant tissue, the time of homogenization can be properly prolonged.)
 - (3) Ultrasonication: Treat the cells with ultrasonic cell disruptor (200 W, 2 s/ time, interval for 3 s, the total time is 5 min).

Appendix III References

1. Neill S J, Desikan R, Clarke A, et al. Hydrogen peroxide and nitric oxide as signalling molecules in plants. *Journal of Experimental Botany*, 2002, 53(372): 1237-1247.
2. Veal E A, Day A M, Morgan B A. Hydrogen peroxide sensing and signaling. *Molecular Cell*, 2007, 26(1): 1-14.
3. Marinho H S, Real C, Cyrne L, et al. Hydrogen peroxide sensing, signaling and regulation of transcription factors. *Redox Biology*, 2014, 2(2): 535-562.
4. Carnevale R, Nocella C, Pignatelli P, et al. Blood hydrogen peroxide breakdown activity in healthy subjects and in patients at risk of cardiovascular events. *Atherosclerosis*, 2018, 274: 29-34.
5. Moloney J N, Cotter T G. ROS signalling in the biology of cancer. *Seminars in Cell & Developmental Biology*, 2018, 80: 50-64.