

## NB110-89717 Protocol

### Immunohistochemistry Protocol specific for Ki67 Antibody (NB110-89717)

[[URL:[https://www.novusbio.com/products/ki67-mki67-antibody\\_nb110-89717](https://www.novusbio.com/products/ki67-mki67-antibody_nb110-89717)]][[Caption:Ki67 Antibody ]]  
Antigen Unmasking:

Bring slides to a boil in 10 mM sodium citrate buffer pH 6.0 then maintain at a sub-boiling temperature for 10 minutes. Cool slides on bench top for 30 minutes.

#### Staining:

- 1) Wash sections in dH<sub>2</sub>O three times for 5 minutes each.
- 2) Wash section in wash buffer (1X PBS/0.1% Tween-20 (1X PBST)) for 5 minutes.
- 3) Block each section with 100-400 ul blocking solution (1X PBST, 5% goat serum) for 1 hour at room temperature.
- 4) Remove blocking solution and add 100-400 ul primary antibody diluted in 1X PBST, 5% goat serum to each section. Incubate overnight at 4C.
- 5) Remove antibody solution and wash sections in wash buffer three times for 5 minutes each.
- 6) Add 100-400 ul biotinylated secondary antibody, diluted in 1X PBST, 5% goat serum. Incubate 30 minutes at room temperature.
- 7) Remove secondary antibody solution and wash sections three times with wash buffer for 5 minutes each.
- 8) Add 100-400 ul Streptavidin-HRP reagent to each section and incubate for 30 minutes at room temperature.
- 9) Wash sections three times in wash buffer for 5 minutes each.
- 10) Add 100-400 ul DAB substrate to each section and monitor staining closely.
- 11) As soon as the sections develop, immerse slides in dH<sub>2</sub>O.
- 12) Counterstain sections in hematoxylin.
- 13) Wash sections in dH<sub>2</sub>O two times for 5 minutes each.
- 14) Dehydrate sections.
- 15) Mount coverslips.