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## **NLS1663 Protocol**

## Immunohistochemistry protocol for EMR2 Antibody (NLS1663)

Ilmmunohistochemistry Protocol for EMR2 Antibody (NLS1663): https://www.novusbio.com/products/emr2-antibody\_nls1663 mmunohistochemistry

- 1. Prepare tissue with formalin fixation and by embedding it in paraffin wax.
- 2. Make 4-um sections and place on pre-cleaned and charged microscope slides.
- 3. Heat in a tissue-drying oven for 45 minutes at 60 degrees Celcius.
- 4. Deparaffinize the tissues by wash drying the slides in 3 changes of xylene approximately 5 minutes each @ RT.
- 5. Rehydrate the tissues by washing the slides in 3 changes of 100% alcohol approximately 3 minutes each @ RT.
- 6. Wash the slides in 2 changes of 95% alcohol approximately 3 minutes each @ RT.
- 7. Wash the slides in 1 change of 80% alcohol approximately 3 minutes @ RT.
- 8. Rinse the slides in gentle running distilled water approximately 5 minutes @ RT.
- 9. Perform antigen retrieval by steaming the slides in 0.01M sodium citrate buffer (pH 6.0) @ 99-100 degrees Celcius for 20 minutes.
- 10. Remove the slides from the heat and let stand in buffer @ RT for 20 minutes.
- 11. Rinse the slides in 1X TBS-T for 1 minute @ RT.

- 12. Begin the immunostaining by applying a universal protein block approximately 20 minutes @ RT.
- 13. Drain protein block from the slides and apply the diluted primary antibody approximately 45 minutes @ RT.
- 14. Rinse the slide in 1X TBS-T approximately 1 minute @ RT.
- 15. Apply a biotinylated anti-rabbit IgG (H+L) secondary approximately 30 minutes @ RT.
- 16. Rinse the slide in 1X TBS-T approximately 1 minute at RT.
- 17. Apply an alkaline phosphatase steptavidin approximately 30 minutes at RT.
- 18. Rinse the slide in 1X TBS-T approximately 1 minute at RT.
- 19. Apply an alkaline phosphatase chromagen substrate approximately 30 minutes at RT.
- 20. Rinse the slide in distilled water approximately 1 minute @ RT.

- 21. Dehydrate the tissue by washing the slides in 2 changes of 80% alcohol approximately 1 minute each @ RT.
- 22. Wash the slides in 2 changes of 95% alcohol approximately 1 minute each @ RT.
- 23. Wash the slides in 3 changes of 100% alcohol approximately 1 minute each @ RT.
- 24. Wash the slides in 3 changes of xyleneapproximately 1 minute each @ RT.
- 25. Apply cover slip.

<sup>\*\*</sup>Do not allow the tissues to dry at any time during the staining procedure\*\*

<sup>\*\*</sup>This method should only be used if the chromagen substrate is alcohol insoluble (ie: Vector Red, DAB)\*\*